

Biological Forum – An International Journal

15(11): 243-248(2023)

ISSN No. (Print): 0975-1130 ISSN No. (Online): 2249-3239

# Screening of BC<sub>3</sub>F<sub>2</sub> Rice population for Submergence Tolerance During Seed Germination

Salomi R.<sup>1</sup>, Vignesh P.<sup>1</sup> and Bharathkumar S.<sup>2\*</sup> <sup>1</sup>Ph.D. Scholar, PG & Research Department of Botany, Kandaswami Kandar's College, Velur-638 182, Namakkal (Tamil Nadu), India. <sup>2</sup>Assistant Professor, PG & Research Department of Botany, Kandaswami Kandar's College, Velur-638 182, Namakkal (Tamil Nadu), India.

(Corresponding author: Bharathkumar S.\*)

(Received: 09 September 2023; Revised: 09 October 2023; Accepted: 19 October 2023; Published: 15 November 2023) (Published by Research Trend)

ABSTRACT: Rice (Oryza sativa L.) is an important food crop in the Asian countries, but it is severely affected by submergence stress from seed germination to reproductive stage during heavy monsoon period every year in some parts of the world. This situation has created to improve highly preferable rice varieties for submergence stress tolerance. In the present study, two short duration rice varieties, ADT36 and ADT37 were improved for submergence tolerance at seedling stage using CR Dhan 801 with Submergence 1 (Sub1) locus as donor up to BC<sub>3</sub>F<sub>2</sub> generation through marker assisted backcross method. In Tamil Nadu state, these two rice varieties are adapted in the Cauvery delta region wherever submergence and drought stress is erratic. Seeds from this generation were evaluated for submergence tolerance at seed germination stage. At parental level, we noted less and more elongation in the coleoptile growth of donor and both recipient parents under anaerobic condition, respectively when compare to aerobic condition. In the evaluation of BC<sub>3</sub>F<sub>2</sub> population, we found a range in the elongation of coleoptile and root length under anaerobic condition. From these, a number of seven rice lines (ADT36- F12-5-19.6; ADT36- F12-13-7.11,13,14, ADT36- F14-5-3.4, ADT36- F14-16-2.6, ADT36- F15-2-5.1) for ADT36 and four rice lines (ADT37- F113-27-5.2, ADT37- F115-2-1.1, ADT37- F115-2-2.5, ADT37- F115-2-2.7) for ADT37 rice variety were found to be superior in coleoptile elongation compared to donor line. These rice lines at seed germination stage followed quiescence strategy like donor parent having Sub1 associated with seedling stage tolerance and they possessed decreased growth under submergence condition. Further, other lines exhibited escaping mechanism under flooding with higher rate of CL and RL. Thus, selected rice lines harboring Sub1 locus linked with submergence tolerance at seedling stage will support for flood tolerance at seed germination stage also in the genetic background of ADT36 and ADT37. Therefore, in future, these lines can be used in both methods of direct seed sowing and transplanting in upland and lowland rice cultivating areas and as a genetic source in the rice breeding program.

Keywords: Submergence tolerance, anaerobic seed germination, Cauvery delta areas, ADT36, ADT37, CR Dhan 801. Coleoptile and Leaf elongation.

### **INTRODUCTION**

Rice is a semi-aquatic crop and it has some adaptive mechanisms to survive and grow under water logged and/or submerged condition for a few days. In rice, development of aerenchyma cells allow sufficient amount of oxygen to submerged parts of the plant during stress. If submergence is prolonged, rice plant loses its survival rate gradually by loss of soil nutrients through reduction of soil redox potential, volatilization and deep percolation (Anjani Kumar et al., 2021). Thus, submergence stress causes yield loss over 600 million to 1 billion US dollars annually only in Asian countries (Dey and Upadhaya 1996) through devastating nearly 22 million ha rice areas (Sarkar et al., 2006). This yield loss is likely to increase in the flood susceptible rice producing areas in the era of global warming (IPCC, 2007). According to previous reports, rice plants are

very sensitive to submergence stress during seed germination and seedling stage (Ismail et al., 2009; Angaji et al., 2010, Joshi et al., 2013). In this connection, identification of AG and SUB1 QTL from rice genotypes supports the rice plants for the higher survival rate during seed germination and seedling stage, respectively. Rice varieties incorporated with SUB1 QTL enhanced their survival rate more during seedling stage through maintaining high and low level of alcohol dehydrogenase activity and chlorophyll degradation rate under submergence, respectively (Anjani Kumar et al., 2021). Rice genotypes with AG QTL overcome the flooded condition under direct seeded rice (DSR) conditions (Ismail et al., 2009, Miro et al., 2017, Chamara et al., 2018, Lal et al., 2018). In Tamil Nadu state, rice cultivation is highly practiced in the Cauvery delta region (CDR) for three times in a year depends on seasonal rains. However, this region is 243

Salomi et al..

Biological Forum – An International Journal 15(11): 243-248(2023)

susceptible to floodling due to heavy rains at the time of monsoon and the rice farmers in this region frequently face the economic cries by the loss of crop, labour and season. In the CDR, many short duration rice varieties including ADT36 and ADT37 are widely cultivated. In the present study, these varieties were improved for submergence tolerance at seedling stage up to  $BC_3F_2$  generation and these lines were evaluated for submergence tolerance during seed germination also under flooding.

#### MATERIALS AND METHODS

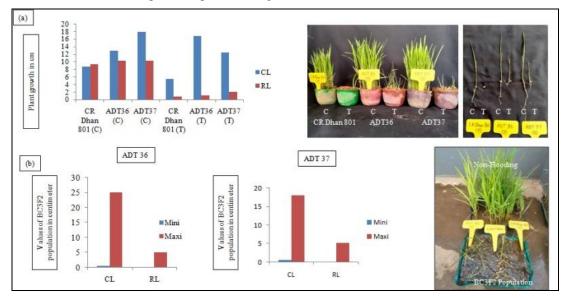
**Rice seeds:** A small quantity of rice seeds of ADT36 and ADT37 from Tamilnadu Rice Research Institute (TRRI), Aduthurai, Tamilnadu state, INDIA and CR Dhan 801 rice seeds from National Rice Research Institute (NRRI), Cuttack, Odisha state were sourced. Here, ADT36 and ADT37 rice varieties were used as female parent (recipient) and CR Dhan801 as male parent (donor) which harboring Submergence 1 (Sub1) locus.

**Production of BC**<sub>3</sub>**F**<sub>2</sub> **population:** F1 seeds were derived from a cross between ADT36 x CR dhan 801(cross-1) and ADT37 x CR dhan 801 (cross-2) and they were subjected to PCR screening using Sub1 gene specific marker Sub1C173 (Septiningsih *et al.* (2009) and identified positive F1 plants with heterozygous allele. Thus, identified plants were backcrossed with ADT36 and ADT37 as recurrent parent (RP) and produced BC<sub>1</sub>F<sub>1</sub>, BC<sub>2</sub>F<sub>1</sub> and BC<sub>3</sub>F<sub>1</sub> generation subsequently based on morphological changes. Finally they were allowed for self-pollination to produce BC<sub>3</sub>F<sub>2</sub> seeds.

Evaluation of BC<sub>3</sub>F<sub>2</sub> population under submergence condition at seed germination stage: Prior to seed sowing, seed dormancy was broken by incubating the seeds at 50°C for up to 5 days. For submergence screening during seed germination, seeds of ADT36, ADT37, CR Dhan 801 and BC<sub>3</sub>F<sub>2</sub> seeds were sowed at 1.0 cm below the soil surface in plastic cups containing fine soil brought from rice field and seeds were covered with fine soil powder. They were placed into a large plastic container and immediately flooded the container carefully and the water level was maintained at 10 cm for 21 days. Similarly, another set up was maintained in non-flooded condition (Ismail *et al.*, 2009). Length of coleoptile and root in parental and BC<sub>3</sub>F<sub>2</sub> lines were measured in both flood and non-flood conditions.

#### **RESULTS AND DISCUSSION**

Submergence/flooding restrict the diffusion of O2 and CO<sub>2</sub> into water and it creates a hypoxic condition (< 21% - O<sub>2</sub>) to plants through making these gases unavailable to plant tissues. Hence, the respiration process of plants in the absence of O<sub>2</sub> is affected and leads to accumulation of phytotoxic substances such as Fe<sup>2+</sup>, Mn<sup>2+</sup>, H<sub>2</sub>S, O<sub>2</sub> radicals and the products of fermentation which cause damages to the plant tissue (Drew and Lynch 1980). Generally, rice seeds germinate under moisture condition and develop the root and shoot system. Interestingly, rice seeds have a capability to germinate and grow under complete submerged condition using an adaptive system called SNORKEL 1 and SNORKEL 2 (SK1/2) (Bailey-Serres et al., 2012a). Hence, rice exempts from other food crops such as wheat (Alpi and Beevers 1983, Perata et al. 1997), oat (Alpi and Beevers 1983), and barley (Perata et al., 1997). It is an escaping strategy and it supports the germinating seeds to tolerate submergence stress (Bailey-Serres et al., 2012). In this study, we evaluated BC<sub>3</sub>F<sub>2</sub> progenies for submergence tolerance at seed germination stage and its results are given in Table 1. Under flooding, elongation of CL and RL was 16.9 and 1.1cm in ADT36, 12.5 and 2.0 cm in ADT37 and 5.5 and 0.76 cm in CR dhan801 respectively. Under non-flooding condition, plant height and root length were 13.0 and 10.3 cm in ADT36, 18.0 and 10.3 cm in ADT37, 8.8 and 9.4 cm in CR dhan 801 respectively (Fig. 1a, b; Table 1).



**Fig. 1.** Plant growth of (a) parental lines and (b) BC<sub>3</sub>F<sub>2</sub> population of ADT36 and ADT37. T-Treatment (Flooding); C-Control (Non-Flooding); CL-Coleoptile length; RL-Root length.

ADT36 x CR dhan 801 cross			ADT37 x CR dhan 801 cross		
Parental lines/BC <sub>3</sub> F <sub>2</sub>	CL	RL	Parental lines/BC <sub>3</sub> F <sub>2</sub>	CL	RL
progenies	(cm)	(cm)	progenies	(cm)	(cm)
CR Dhan 801 (C)	8.9	9.2	CR Dhan 801 (C)	8.8	9.4
ADT36 (C)	13.0	10.26	ADT37 (C)	18.0	10.30
CR Dhan 801 (T)	5.5	0.76	CR Dhan 801 (T)	5.5	0.76
ADT36 (T)	16.9	1.1	ADT37 (T)	12.5	2.0
ADT36- F12-5-19.1	16.0	0.3	ADT37- F <sub>1</sub> 13-24-3.7	13.5	0.3
ADT36- F12-5-19.6	5.0	0.2	ADT37- F113-24-3.12	12.0	0.2
ADT36- F12-5-19.7	15.0	1.5	ADT37- F113-24-3.13	11.0	1.5
ADT36- F12-5-19.9	9.0	1.2	ADT37- F <sub>1</sub> 13-24-3.14	9.0	1.2
ADT36- F <sub>1</sub> 2-5-19.10	8.0	0.3	ADT37- F <sub>1</sub> 13-24-3.15	11.8	0.3
ADT36- F12-5-19.13	10.5	0.5	ADT37- F <sub>1</sub> 13-27-5.1	10.5	0.5
ADT36- F <sub>1</sub> 2-5-19.14	16.0	2.2	ADT37- F <sub>1</sub> 13-27-5.2	4.0	0.2
ADT36- F12-5-19.15	25.0	4.0	ADT37- F <sub>1</sub> 13-27-5.3	8.0	4.0
ADT36- F <sub>1</sub> 2-5-19.16	16.5	0.5	ADT37- F <sub>1</sub> 13-27-5.5	7.0	0.5
ADT36- F12-5-19.17	16.5	5.0	ADT37- F <sub>1</sub> 13-27-5.6	8.2	5.0
ADT36- F <sub>1</sub> 2-5-19.18	7.0	0.5	ADT37- F <sub>1</sub> 13-27-5.10	9.0	0.5
ADT36- F12-5-19.20	8.5	3.5	ADT37- F <sub>1</sub> 15-2-1.1	0.5	0.0
ADT36- F12-13-7.1	18.0	1.0	ADT37- F115-2-1.7	6.0	1.0
ADT36- F <sub>1</sub> 2-13-7.2	5.5	0.6	ADT37- F <sub>1</sub> 15-2-1.7	5.5	0.6
ADT36- F12-13-7.3	7.5	0.0	ADT37- F115-2-1.10	7.5	0.0
	12.0			18.0	1.0
ADT36- F12-13-7.4	8.0	1.0 0.2	ADT37- F <sub>1</sub> 15-2-1.13	8.0	0.2
ADT36- F <sub>1</sub> 2-13-7.5 ADT36- F <sub>1</sub> 2-13-7.6	8.0	0.2	ADT37- F <sub>1</sub> 15-2-1.14 ADT37- F <sub>1</sub> 15-2-1.16	7.5	0.2
	6.5	0.2	ADT37- F115-2-2.1	16.5	0.2
ADT36- F12-13-7.9	17.5	0.2			0.2
ADT36- F12-13-7.10			ADT37- F <sub>1</sub> 15-2-2.2	13.5	
ADT36- F12-13-7.11	5.0	0.0	ADT37- F115-2-2.4	15.0	0.0
ADT36- F12-13-7.12	8.5	1.0	ADT37- F115-2-2.5	0.5	0.0
ADT36- F12-13-7.13	4.0	0.0	ADT37- F115-2-2.7	5.0	0.0
ADT36- F12-13-7.14	5.0	0.5	ADT37- F115-2-2.10	9.0	1.5
ADT36- F12-13-7.15	10.0	3.0	ADT37- F115-2-2.14	7.0	3.0
ADT36- F <sub>1</sub> 2-13-7.16	17.2	1.0	ADT37- F <sub>1</sub> 15-2-2.18	12.2	1.0
ADT36- F <sub>1</sub> 2-13-7.17	18.0	0.5			
ADT36- F <sub>1</sub> 2-13-7.18	9.0	0.3			
ADT36- F <sub>1</sub> 2-13-7.19	0.6	0.0			
ADT36- F <sub>1</sub> 2-13-7.20	5.5	3.0			
ADT36- F14-5-3.4	5.0	0.2			
ADT36- F14-5-3.5	8.5	0.0			
ADT36- F14-5-3.7	11.7	0.5			
ADT36- F14-5-3.8	10.5	0.0		┼───┤	
ADT36- F <sub>1</sub> 4-5-3.9	13.2	1.5		┨────┤	
ADT36- F14-5-3.10	11.0	0.5		┨────┤	
ADT36- F14-16-2.1	0.5	0.0			
ADT36- F14-16-2.3	13.0	1.5			
ADT36- F14-16-2.6	5.0	0.5			
ADT36- F14-16-2.7	11.0	0.5			
ADT36- F14-16-2.10	10.5	0.5			
ADT36- F <sub>1</sub> 5-2-5.1	4.5	0.5			
ADT36- F15-2-5.2	17.3	2.5			
ADT36- F15-2-5.7	9.4	3.0			
ADT36- F15-2-5.9	9.0	2.5			
ADT36- F <sub>1</sub> 5-2-5.17	10.5	0.5			
ADT36- F18-2-5.8	13.1	1.5			
ADT36- F <sub>1</sub> 8-2-5.25	7.8	2.0			

#### Table 1: Response of parental and BC<sub>3</sub>F<sub>2</sub> lines under anaerobic condition during seed germination.

C – Control (Non-Flooding); T – Treatment (Flooding); CL – Coleoptile length; RL – Root length.

In ADT36 and ADT37, the growth rate of CL is increased to 76.9% and 30.5% and RL is decreased to 89.3% and 80.6% under flooding, respectively. In CR dhan801, elongation of CL is reduced to 37.5% and RL to 91.9% under flooding. At parental level, growth rate of CL was significantly higher when compare to RL under flooding compared to non-flooding condition. Notably, elongation of CL in ADT36 and ADT37 with *Salomi et al.*, *Biological Forum – An International*.

no Sub1 was found to be increased to 67.4% and 30.5% respectively than that of CR Dhan 801 with Sub1 under flooding. This coleoptile elongation is dependent on mobilization rate of stored starch in rice seed during seed germination by the activity of  $\alpha$ -amylase ( $\alpha$ Amy) (1,4- $\alpha$ -D-glucan maltohydrolase). During the mobilization,  $\alpha$ -1,4-glucosidic bonds of starch is hydrolyzed to yield  $\alpha$ -glucose and  $\alpha$ -maltose (Damaris

et al., Biological Forum – An International Journal 15(11): 243-248(2023)

et al., 2019, Senapati et al., 2019). So far, three subfamilies of aAmy genes such as RAmy1 (A, B, and C), RAmy2A, and RAmy3 (A, B, C, D, E, and F) have been identified (Huang et al., 1990, 1992) and particularly, RAmy3D enzyme is reported for its significant role in the fermentative metabolism pathway for the production of energy under submergence (Hwang et al., 1990). Moreover, the rate of coleoptile elongation is determined by the pre-formed cells in embryo (Jones and Rost 1989) and some phytohormones like ethylene play a significant role in the mechanism in the growth of submerged rice coleoptile (Masuda et al., 1998). Moreover, the rapid coleoptile elongation is linked with decreased and increased level of abscisic acid (ABA) and gibberellic acid (GA), respectively due to the accumulation of ethylene under submerged condition of rice (Hoffmann-Benning & Kende 1992; Fukao and Bailey-Serres 2008). According to Saika et al., (2007), ABA conversion to an inactive form is catalysed through production of ABA 8'- hydroxylase enzyme by stimulation of ethylene which is trapped during submergence.

In BC<sub>3</sub>F<sub>2</sub> population of cross-2, CL and RL elongation was in a range of 0.5 to 25.0 cm and 0.0 to 5.0 cm, respectively whereas in cross-2, they were in the range of 0.5 to 25.0 cm and 0.0 to 5.0 cm, respectively. In  $BC_3F_2$  population under flooding, we noted that 79.1%, 15.4% and 20.8% of progenies showed lower value than ADT36, ADT37 and CR dhan 801, respectively. Variations in coleoptile elongation under submergence influences seedling establishments by a trait called anaerobic germination (AG) which supports for starch degradation to escape seed germination from submergence stress (Huang et al., 2003). In previous surveys also, genetic factors responsible for AG have showed a wide phenotypic variation in survival rate under flooding (Ismail et al., 2009; Miro and Ismail 2013). Specifically, AG1 gene belonging to the trehalose-6-phosphate phosphatase gene-family (Angaji, 2008; Angaji et al., 2010; Baltazar et al., 2014; Septiningsih et al., 2013) helps to elongate the coleoptile under anaerobic condition (Kretzschmar et al., 2015). Coleoptile elongation in rice under anaerobic condition is linked with some specific expansins such EXPA2, EXPA4, EXPA1, EXPB11 as and EXPB17 (Lasanthi-Kudahettige et al., 2007). In recent study also, it is reported that coleoptile elongation showed difference in flooding (Aung et al., 2023). Based on shoot elongation during seed germination, many efficient rice genotypes are identified for flood-prone areas (Vijay Kumar Reddy et al., 2022).

In the study of root length in F<sub>2</sub> population under flooding, 31.2%, and 39.6% of progenies accounted for more value than ADT36 and CR dhan 801 in cross-1 whereas in cross-2, 11.5% and 10.0% of progenies for more value than ADT37 and CR dhan 801. More and less value of root length development under flooding is dependent on coleoptile elongation which reaches the water surface in order to supply oxygen to the root and endosperm for speedy establishment of seedlings following the starch degradation (Magneschi and Perata Salomi et al.,

2009). Then, shoots and roots develop through the events of splitting, aerenchyma cell formation and senescence in submerged coleoptile in response to air (Kawai and Uchimiya 2000). In a study, coleoptiles elongation regulating a microRNA miR393a during seed germination and seedling establishment under submergence is reported in rice and its overexpression exhibits primary root elongation and inhibits coleoptile elongation. The opposite effects of miR393a on root and coleoptile elongation indicate that auxin signalling negatively regulates primary root elongation but positively regulates coleoptile elongation (Guo et al., 2016). A recent finding reports that pyramiding of GERMINATION ANAEROBIC 1 (AG1) locus TREHALOSE 6-PHOSPHATE encoding PHOSPHATASE 7 (TPP7) and SUMBERGENCE 1 (SUB1) locus encoding ethylene-responsive transcription factor SUB1A-1 slowed the elongation growth in near-isogenic lines (Shin et al., 2022, Alam et al., 2020). Therefore, plant growth in CR Dhan 801 having Sub1 locus is controlled but not in ADT36 and ADT37 rice variety. In this connection, rice variety with Sub1 locus undergoes another strategy called Quiescence (Bailey-Serres et al., 2012) during vegetative stage stress. Expression of SUB1A gene inhibits rice elongation through stimulating alcohol *dehydrogenase* (ADH) gene activation for upregulating the expression of slender rice-1 (SLR1) and SLR1 like*l*(*SLRL1*) which are negative regulators of gibberellins (GA) signaling. Hence, SUB1A gene expression reduces the carbohydrate consumption and increases the survival rate of rice seedlings (Xu et al., 2006, Fukao et al., 2006, Fukao et al., 2008).

## CONCLUSIONS

In the present study, we found the difference between the donor and recipient parent based on low and high of coleoptile elongation under flooding. rate respectively. Shoot elongation under flooding which results in consumption of stored starch content very fast and it leads to plant death or lodging following the desubmergence. According to the previous studies, Sub1 locus is linked with only submergence tolerance at seedling stage and it controls the degradation of starch due to shoot elongation but not at seed germination. Elongation of coleoptile in ADT36 and ADT37 rice variety during seed germination associates with SNORKEL 1 and SNORKEL 2 (SK1/2)-dependent escape strategy and CR Dhan 801 with SUBMERGENCE 1A (SUB1A)-dependent quiescence strategy. However, we found both strategies in  $BC_3F_2$ population. Supportively, in a very recent study, rice line pyramided with AG and SUB1 gene has showed negative impact on shoot elongation. Here also, we found difference in flood tolerance between ADT36 and ADT37 i.e. when compare to ADT36, elongation of coleoptile is found to be decreased and increased under flooding and non-flooding condition, respectively. Thus, this study reveals that expression of gene/QTL depends on the genetic background of a rice variety. Further, the screening process of existing genetic source may lead to a way to some other stress tolerance also.

Biological Forum – An International Journal 15(11): 243-248(2023)

#### FUTURE SCOPE

In this study, we identified some rice lines with controlled plant growth in order to tolerate flooding. Therefore, these rice lines can be used as genetic source in rice breeding and they will be suitable for direct seed sowing method in the upland rice cultivating areas.

Acknowledgement. We sincerely thank Tamilnadu Rice Research Institute (TRRI), Aduthurai, Tamilnadu state, INDIA and National Rice Research Institute (NRRI), Cuttack, Odisha state for providing rice seeds. We thank Kandaswami Kandar's College, Velur, Namakkal (District), Tamil Nadu for providing facilities to carry out this experiment **Conflict of interest.** None.

#### REFERENCES

- Alam, R., Hummel, M., Yeung, E., Locke, A. M., Ignacio, J. C. I. and Baltazar, M. D. (2020). Flood resilience loci Submergence 1 and Anaerobic germination 1 interact in seedlings established underwater. *Plant Direct, 4*, e00240.
- Alpi, S. A. and Beevers, H. (1983). Effects of O<sub>2</sub> concentration on rice seedlings. *Plant Physiology*, 71, 30–34.
- Angaji, S. A. (2008). Mapping QTLs for submergence tolerance during germination in rice. *African Journal* of Biotechnology, 7, 2551–2558.
- Angaji, S. A, Septiningsih, E. M., Mackill, D. J. and Ismail, A. M. (2010). QTLs associated with tolerance of flooding during germination in rice (*Oryza* sativa L.). Euphytica, 172, 159–168.
- Anjani Kumar, A. K., Nayak, P. S., Hanjagi, Kavita Kumari, Vijayakumar S, Sangita Mohanty, Rahul Tripathi and Panneerselvam, P. (2021). Submergence stress in rice: Adaptive mechanisms, coping strategies and future research needs. *Environmental and Experimental Botany*, 186, 104448.
- Aung, K. M., Oo, W. H., Maung, T. Z., Min, M. H., Somsri, A., Nam, J., Kim, K. W., Nawade, B., Lee, C. Y., Chu, S. H. and Park, Y. J. (2023). Genomic landscape of the *OsTPP7* gene in its haplotype diversity and association with anaerobic germination tolerance in rice. *Front Plant Sciences*, 14, 1225445.
- Bailey-Serres, J., Fukao, T., Gibbs, D. J., Holdsworth, M. J., Lee, S. C., Licausi, F. and van Dongen, J. T. (2012a). Making sense of low oxygen sensing. *Trends in Plant Science*, 17, 129–138.
- Bailey-Serres, J., Lee, S. C. and Brinton, E. (2012). Waterproofing crops: Effective flooding survival strategies. *Plant Physiology*, 160, 1698–1709.
- Baltazar, M. D., Ignacio, J. C. I., Thomson, M. J., Ismail, A. M., Mendioro, M.S. and Septiningsih, E. M. (2014). QTL mapping for tolerance of anaerobic germination from IR64 and the aus landrace Nanhi using SNP genotyping. *Euphytica*, 197, 251–260.
- Chamara, B. S., Marambe, B., Kumar, V., Ismail, A. M., Septiningsih, E. M. and Chauhan, B. S. (2018). Optimizing sowing and flooding depth for anaerobic germination-tolerant genotypes to enhance crop establishment, early growth, and weed management in dry-seeded rice (*Oryza sativa* L.). *Frontiers in Plant Science*, 9,
- Damaris, R. N., Lin, Z., Yang, P. and He, D. (2019). The Rice Alpha-Amylase, Conserved Regulator of Seed Maturation and Germination. *International Journal of Molecular Sciences*, 20, 450.

- Drew, M. and Lynch, J. (1980). Soil Anaerobiosis, Microorganisms, and Root Function. Annu. Rev. Phytopathol. 18, 37–66.
- Dey, M. M. and Upadhyaya H. K. (1996). Yield loss due to drought, cold and submergence in Asia. In: Evenson R, Herdt R, Hossain M (eds) Rice research in Asia: progress, priorities. CAB International, Wallingford, pp 291–303
- Fukao, T. and Bailey-Serres, J. (2008). Submergence tolerance conferred by Sub1A is mediated by SLR1 and SLRL1 restriction of gibberellin responses in rice. *Proceedings of the National Academy of Sciences*, 105, 16814–16819.
- Fukao, T., Xu, K., Ronald, P. C. and Bailey-Serres, J. (2006). A variable cluster of ethylene response factor-like genes regulates metabolic and developmental acclimation responses to submergence in rice. *Plant Cell*, 18, 2021–2034.
- Guo, F., Han, N., Xie, Y., Fang, K., Yang, Y., Zhu, M., Wang, J. and Bian, H. (2016). The miR393a/target module regulates seed germination and seedling establishment under submergence in rice (*Oryza sativa* L.). *Plant, Cell and Environment*, 39, 2288–2302.
- Hattori, Y., Nagai, K., Furukawa, S., Song, X. J., Kawano, R., Sakakibara, H., Wu, J., Matsumoto, T., Yoshimura, A. and Kitano, H. (2009). The ethylene response factors SNORKEL1 and SNORKEL2 allow rice to adapt to deep water. *Nature*, 460, 1026–1030.
- Hoffmann-Benning, S. and Kende, H. (1992). On the role of abscisic acid and gibberellin in the regulation of growth in rice. *Plant Physiology*, 99, 1156–1161.
- Huang, N., Stebbins, G. and Rodriguez, R. (1992). Classification and evolution of α-amylase genes in plants. *Proceedings of the National Academy of Sciences*, 89, 7526–7530.
- Huang, N., Sutliff, T. D., Litts, J. C. and Rodriguez, R. L. (1990). Classification and characterization of the rice alpha-amylase multigene family. *Plant Molecular Biology*, 14, 655–668.
- Hwang, Y. S., Thomas, B. R. and Rodriguez, R. L. (1990). Differential expression of rice α-amylase genes during seedling development under anoxia. *Plant Molecular Biology*, 40, 911–920.
- IPCC. (2007). Intergovernmental Panel on Climate Change (IPCC): The Physical Science Basis (eds Solomon, S. et al.) (Cambridge Univ. Press, 2007).
- Ismail, A. M., Ella, E. S., Vergara, G. V. and Mackill, D. J. (2009). Mechanisms associated with tolerance to flooding during germination and early seedling growth in rice (*Oryza sativa*). *Annals of Botany*, 103, 197– 209.
- Jones, T. J. and Rost, T. L. (1989). The developmental anatomy and ultrastructure of somatic embryos from rice (*Oryza sativa* L.) scutellum epithelial-cells. *Botanical Gazette*, 150, 41–49.
- Joshi, E., Kumar, D., Lal, B., Nepalia, V., Gautam, P., Vyas, A.K., (2013). Management of direct seeded rice for enhanced resource - use efficiency. *Plant Knowledge Journal*, 2, 119–134.
- Kawai, M. and Uchimiya, H. (2000). Coleoptile senescence in rice (*Oryza sativa* L.) *Annals of Botany*, 86, 405–414.
- Kretzschmar, T., Pelayo, M. A., Trijatmiko, K. R., Gabunada, L. F., Alam, R., Jimenez, R., Mendioro, M. S., Slamet-Loedin, I. H., Sreenivasulu, N. and Bailey-Serres, J. (2015). A trehalose-6-phosphate phosphatase enhances anaerobic germination tolerance in rice. *Nature Plants*, 1, 15124.

Salomi et al.,

- Lal, B., Gautam, P., Nayak, A. K., Raja, R., Shahid, M., Tripathi, R., Singh, S., Septiningsih, E. M. and Ismail, A. M. (2018). Agronomic manipulations can enhance the productivity of anaerobic tolerant rice sown in flooded soils in rainfed areas, *Field Crops Research*, 220, 105-116.
- Lasanthi-Kudahettige, R., Magneschi, L., Loreti, E., Gonzali, S., Licausi, F., Novi, G., Beretta, O., Vitulli, F., Alpi, A. and Perata, P. (2007). Transcript Profiling of the Anoxic Rice Coleoptile. *Plant Physiology*, 144, 218– 231.
- Magneschi, L. and Perata, P. (2009). Rice germination and seedling growth in the absence of oxygen. Annals of Botany, 103, 181–196.
- Masuda, Y., Kamisaka, S. and Hoson, T. (1998). Growth behaviour of rice coleoptiles. *Journal of Plant Physiology*, 152, 180–188.
- Miro, B. and Ismail, A. M. (2013). Tolerance of anaerobic conditions caused by flooding during germination and early growth in rice (*Oryza sativa* L.). Front Plant Sciences, 4, 269.
- Perata, P., Guglielminetti, L. and Alpi, A. (1997). Mobilization of endosperm reserves in cereal seeds under anoxia. *Annals of Botany*, 79, 49–56.
- Saika, H., Okamoto, M., Miyoshi, K., Kushiro, T., Shinoda, S., Jikumaru, Y. and Nakazono, M. (2007). Ethylene promotes submergence-induced expression of OsABA80x1, a gene that encodes ABA 8'hydroxylase in rice. *Plant Cell Physiology*, 26, 287– 298.
- Sarkar, R. K., Reddy, J. N., Sharma, S. G. and Ismail, A. M. (2006). Physiological basis of submergence tolerance in rice and implications for crop improvement. *Current Sciences*, 91, 899–906.

- Senapati, S., Kuanar, S. and Sarkar, R. (2019). Anaerobic Germination Potential in Rice (*Oryza sativa L.*): Role of Amylases, Alcohol deydrogenase and Ethylene. *Journal of Stress Physiology and Biochemistry*, 15, 39–52.
- Septiningsih, E. M., Ignacio, J. C., Sendon, P. M., Sanchez, D. L., Ismail, A. M. and Mackill, D. J. (2013). QTL mapping and confirmation for tolerance of anaerobic conditions during germination derived from the rice landrace Ma-Zhan Red. *Theoretical Applied Genetics*, 126, 1357–1366.
- Septiningsih, E. M., Pamplona, A. M., Sanchez, D. L., Neeraja, C. N., Vergara, G. V., Heuer, S., Ismail, A. M. and Mackill, D. J. (2009). Development of submergence-tolerant rice cultivars: the Sub1 locus and beyond. *Annals in Botany*, 103, 151-160.
- Shin, N. H., Han, J. H., Vo, K. T. X., Seo, J., Navea, I. P., Yoo, S. C., Jeon, J. S. and Chin, J. H. (2022). Development of a Temperate Climate-Adapted indica Multi-stress Tolerant Rice Variety by Pyramiding Quantitative Trait Loci. *Rice* (N Y), 15(1), 22.
- Vijay Kumar Reddy, C., Pradhan, B., Anandan, A., Manasi Dash, Samal, K. C. and Panda, R. K. (2022). Screening of rice genotypes for anaerobic germination tolerance: Identifying potential breeding lines. *Biological Forum-An International Journal*, 14(4a), 687-693.
- Xu, K., Xu, X., Fukao, T., Canlas, P., Maghirang-Rodriguez, R., Heuer, S., Ismail, A. M., Bailey-Serres, J., Ronald, P. C. and Mackill, D. J. (2006). Sub1A is an ethyleneresponse-factor-like gene that confers submergence tolerance to rice. *Nature*, 442, 705–708.

**How to cite this article:** Salomi R., Vignesh P. and Bharathkumar S. (2023). Screening of BC<sub>3</sub>F<sub>2</sub> Rice population for Submergence Tolerance During Seed Germination. *Biological Forum – An International Journal*, *15*(11): 243-248.