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# Study of Simple Measures of Genetic Variability Parameters in F<sub>2:3</sub> Populations of Cowpea (*Vigna unguiculata* L. Walp.)

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ABSTRACT: Two F<sub>2:3</sub> populations of cowpea were evaluated for 10 different quantitative traits during *summer* 2021 to study genetic variability parameters. Analysis of variance revealed significant differences among all the segregants of the population for most of the traits studied. High phenotypic coefficient of variance and genetic coefficient of variance were recorded for plant height, number of primary branches/plant, dry matter/plant and seed yield/plant in F<sub>2:3</sub> population of VBN-1 × RC-19; number of primary branches/plant, number of pods/plant, dry matter/plant and seed yield/plant in F<sub>2:3</sub> population of KBC-9 × PGCP-6. High heritability and GAM were recorded for plant height, number of primary branches/plant, number of pods/plant, pod length, number of seeds/pod, dry matter/plant and seed yield/plant in the F<sub>2:3</sub> population of VBN-1 × RC-19; plant height, number of primary branches/plant, number of seeds/pod, dry matter/plant and seed yield/plant in the F<sub>2:3</sub> population of VBN-1 × RC-19; plant height, number of primary branches/plant, number of seeds/pod, dry matter/plant and seed yield/plant in the F<sub>2:3</sub> population of VBN-1 × RC-19; plant height, number of primary branches/plant, number of seeds/pod, dry matter/plant and seed yield/plant in the F<sub>2:3</sub> population of VBN-1 × RC-19; plant height, number of primary branches/plant, number of seeds/pod, dry matter/plant and seed yield/plant. This indicates the scope for improvement of these characters through selection.

Keywords: Cowpea, genetic coefficient of variance, phenotypic coefficient of variance, heritability, variability.

#### **INTRODUCTION**

Cowpea (*Vigna unguiculata* L. Walp.) is a warmseason, annual, self-pollinated diploid grain legume with a 620 Mb genome size (Munoz *et al.*, 2017). It is commonly known as Lobia. Cowpea has been named so because of its use as cattle feed. It is commonly known as Lobia. It is also known by its different vernacular names *viz.*, Rawan (Hindi), Chavali (Marathi) and Barbate (Bengali). Some of the cowpea cultivars grown for their immature pods or vegetable purpose are known as Asparagaus bean, snake bean or yard long bean and when they are grown for dry seeds, it is also known as black eyed pea, kaffir pea and southern pea.

Its dry edible grains are rich in protein (20-32%) with high amounts of essential amino acids (lysine and tryptophan), minerals (zinc, iron, calcium), vitamins (thiamine, folic acid and riboflavin) and fibre (6%) with low fat (<1%) (Sebetha *et al.*, 2014; Boukar *et al.*, 2015). Cowpea is a multifunctional legume grown for food, fodder, vegetables and green manure (Timko and Singh, 2008; Gonçalves *et al.*, 2016). Cowpea can be cultivated in subtropical, semitropical, and desert environments. It grows well on poor soil with a wide range of soil pH and it is resistant to high temperatures and low water constraints, thus it is an excellent choice for resource-limited small-scale farmers to sustenance (Carvalho *et al.*, 2017). The development of an effective crop improvement programme depends upon the existence of genetic variability. Genetic variability is the measure of the tendency of distinct genotypes in a population to vary from each other. Variability depends on genetic factors, environmental factors (edaphic and climatic), and bioactive compounds (caused by physiological factors). The degree to which the variability of a quantitative character is transmitted to the progeny is referred to as heritability. It provides valuable biometrical concepts and has been considered to be an index of the effectiveness of selection because it helps in proportioning the total variation into heritable and environmental effects (Johnson *et al.*, 1955; Khan *et al.*, 2015).

## MATERIALS AND METHODS

The experimental material consisted of 100 and 70  $F_3$  family rows derived from crosses VBN-1 × RC-19 (cross I) and KBC-9 × PGCP-6 (cross II), respectively. These  $F_3$  family rows were evaluated during *summer* 2021 using Augmented Block Design with five checks (C-152, KBC-2, KBC-9, PGCP-6 and IT-803695-1) replicated five times and parents at the Department of Genetics and Plant Breeding, College of Agriculture, Kalaburagi. Each  $F_3$  progeny family was sown in rows with a length of 4 meters and a spacing of 45 cm × 10 cm. Five competitive plants selected randomly from

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each family row were used to record observation on ten traits *viz.*, days to initiation of flowering, days to physiological maturity, plant height (cm), number of primary branches/plant, number of pods/plant, number of seeds/pod, pod length, test weight (g), dry matter/plant (g), seed yield/plant (g). The mean data of all traits were recorded and utilized for statistical analysis viz., genetic coefficient of variation (%), phenotypic coefficient of variation (%), heritability (%) (broad sense), genetic advance and genetic advance as per cent mean (%).

## **RESULTS AND DISCUSSION**

Analysis of variance revealed that most of the segregants recorded significant variation for most of the traits, and it indicated the presence of sufficient variability for these characters (Table 1 and 2). ANOVA revealed that all segregants of both the populations were significantly different for all the traits studied. Similarly, the effects due to block, due to treatment, and due to check and variety were also significant for most of the traits studied, except days to maturity in the  $F_{2:3}$  population of KBC-9 × PGCP-6. This indicated that there is a significantly higher level of variability present among the segregating populations for all the yield-related characters studied. Similar findings were also observed by Verma *et al.* (2019), Nair *et al.* (2018) and Khan *et al.* (2015).

Estimates of variability components viz., mean, range, genotypic coefficient of variation (GCV),

phenotypic coefficient of variation (PCV), heritability ( $h^2$ ) and genetic advance as per cent of mean (GAM) for all the yield contributing traits are presented in Table 3 for F<sub>2:3</sub> population of VBN-1 x RC-19 (Fig. 1 and Fig. 2) and Table 4 for F<sub>2:3</sub> population of KBC-9 x PGCP-6 (Fig. 3 and Fig. 4). Higher phenotypic coefficient of variation values than that of genotypic coefficient of variation values indicated the influence of environment on all traits. But, narrow differences between PCV and GCV values were observed for all the traits.

High PCV and GCV were recorded for plant height (21.06%, 20.46%), number of primary branches/plant (28.27%, 27.23%), dry matter/plant (22.45%, 21.98%) and seed yield/plant (27.47%, 26.66%) in  $F_{2:3}$  population of VBN-1  $\times$  RC-19; number of primary branches/plant (31.89%, 31.18%), number of pods/plant (27.31%, 26.73%), dry matter/plant (24.53%, 23.42%) and seed yield/plant (31.77%, 31.37%) in  $F_{2:3}$  population of KBC-9 × PGCP-6 suggesting that sufficient amount of variability is present among the segregants which can be further used in crop improvement programme. Similar findings were observed by Nair et al. (2018), Sheela (2017), Mahesh et al. (2017), Bhadru and Navale (2012), Khan et al. (2015), Meenatchi et al. (2019), Sabale et al. (2018), Shanko et al. (2014) and Khan et al. (2013).

	DF	<b>X</b> 1	$\mathbf{X}_2$	<b>X</b> 3	<b>X</b> 4	X5	X <sub>6</sub>	<b>X</b> 7	X8	X9	X10
Block	4	169.22**	129.83**	258.35**	0.57 **	14.37**	9.55**	11.40 **	1.69 *	104.36 **	49.33 **
Check	6	93.77	95.54*	3.95**	31.74**	485.35**	35.00**	94.40**	161.28**	232.21**	10.62**
Entries	106	90.34**	96.69**	164.56**	1.51**	11.58**	6.92 **	6.44 **	3.05 **	81.88 **	42.03 **
Genotype	99	50.89*	64.07**	154.62**	1.30**	8.10**	5.23 **	5.22 **	1.81 **	74.71 **	37.57 **
Checks vs. Genotype	1	457.14**	3.85	39.39*	0.32*	12.00**	30.89**	22.87 **	70.59 **	184.92 **	20.49 **
Error	24	23.26	16.59	6.67	0.07	1.04	0.82	0.65	0.56	2.36	1.70

Table 1: Analysis of variance in F2:3 segregating generation of cross VBN-1 × RC-19.

Note: \* Significance at 0.05% of probability \*\* Significance at 0.5% of probability

Where,  $X_1$ =Days to initiation of flowering,  $X_2$ =Days to physiological maturity,  $X_3$ =Plant height,  $X_4$ =Number of branches/plant,  $X_5$ = Number of pods/plant,  $X_6$  = Pod length,  $X_7$  = Number of seeds/pod,  $X_8$  =Test weight,  $X_9$  =Total dry matter content/plant,  $X_{10}$  =Seed yield/plant

Table 2: Analysis of variance in F2:3 segregating generation of cross KBC-9 × PGCP-6.

	DF	<b>X</b> 1	<b>X</b> <sub>2</sub>	<b>X</b> 3	<b>X</b> 4	X5	X6	<b>X</b> 7	X8	X9	X10
Block	4	90.42**	208.14*	170.68**	2.02**	12.39**	1.42	5.14**	0.05	125.35**	63.92**
Check	6	45.60*	53.65	0.66**	6.47**	170.03**	5.6 **	35.09**	12.99 **	29.69**	5.29**
Entries	76	52.76**	67.28	114.88**	1.46**	13.80**	2.93**	3.24**	1.43**	62.89**	31.96**
Genotype	69	42.88**	50.69	111.08**	1.57**	11.00**	2.88**	3.08**	0.99**	66.24**	33.55**
Checks vs. Genotype	1	167.94**	512.24**	75.58*	0.01	146.43**	1.73	6.24**	13.27**	71.38**	33.37**
Error	24	14.39	58.47	10.72	0.05	0.32	0.81	0.38	0.41	4.15	0.58

Note: \* Significance at 0.05% of probability \*\* Significance at 0.5% of probability

Where,  $X_1$ =Days to initiation of flowering,  $X_2$ =Days to physiological maturity,  $X_3$ =Plant height,  $X_4$ =Number of branches/plant,  $X_5$ = Number of pods/plant,  $X_6$  = Pod length,  $X_7$  = Number of seeds/pod,  $X_8$ =Test weight,  $X_9$  =Total dry matter content/plant,  $X_{10}$ =Seed yield/plant

Table 3: Estimation of mean and genetic variability parameters in F2:3 segregating generation of cross VBN-1 × RC-19.
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		Mean	Ra	inge	Coefficient	of variation	h <sup>2</sup> (bs)	GAM at 5% mean
Sr. No.	Character		Minimum	Maximum	PCV (%)	GCV (%)	(%)	
1.	Days to initiation of flowering	61.85	49.00	76.00	10.81	7.45	47.48	10.58
2.	Days to physiological maturity	97.86	83.00	116.00	7.42	6.14	68.54	10.48
3.	Plant height (cm)	51.95	19.00	74.00	21.06	20.46	94.41	40.96
4.	Number of branches/plant	3.54	2.00	6.00	28.27	27.23	92.73	54.02
5.	Number of pods/plant	13.63	6.00	18.00	18.53	16.96	83.79	31.99
6.	Pod length (cm)	14.16	9.34	18.84	14.36	12.87	80.31	23.76
7.	Number of seeds/pod	12.58	8.00	17.00	16.08	14.77	84.38	27.95
8.	Test weight (g)	9.67	7.90	14.00	12.87	10.17	62.50	16.58
9.	Total dry matter content/plant (g)	33.59	16.43	54.90	22.45	21.98	95.88	44.35
10.	Seed yield/plant (g)	19.55	8.82	35.21	27.47	26.66	94.14	53.29

			Ra	nge	Coefficient	of variation	h <sup>2</sup> (bs)	GAM
Sr. No.	Character	Mean	Minimum	Maximum	PCV (%)	GCV (%)	(%)	at 5% mean
1	Days to initiation of flowering	67.77	52.00	77.00	8.58	6.52	57.73	10.20
2	Days to physiological maturity	103.45	86.00	117.00	7.02	2.23	10.10	1.46
3	Plant height (cm)	49.19	27.67	73.67	18.19	16.86	86.60	32.32
4	Number of branches/plant	3.28	2.00	6.00	31.89	31.18	95.58	62.79
5	Number of pods/plant	10.00	4.00	17.00	27.31	26.73	95.79	53.89
6	Pod length (cm)	12.56	9.00	17.50	11.94	9.53	63.67	15.66
7	Number of seeds/pod	10.43	7.00	15.00	14.43	13.14	82.91	24.64
8	Test weight (g)	11.25	7.90	14.00	7.94	5.59	49.44	8.09
9	Total dry matter content/plant (g)	28.11	16.62	59.76	24.53	23.42	91.16	46.06
10	Seed yield/plant (g)	15.31	7.21	34.90	31.77	31.37	97.52	63.82

Moderate PCV and GCV were recorded for the number of pods/plant (18.53%, 16.96%), pod length (14.36%, 12.87%), number of seeds/ pod (16.07%, 14.77%) and test weight (12.87%, 10.17%) in  $F_{2:3}$  population of VBN-1 × RC-19; plant height (18.19%, 16.86%) and number of seeds/pod (14.43%, 13.14%) in F<sub>2:3</sub> population of KBC-9 × PGCP-6. Similar findings were also recorded by Sabale et al. (2018) and Dinesh et al. (2017a). Low PCV and GCV were recorded for days to physiological maturity (7.42%, 6.14%) in  $F_{2:3}$ population of VBN-1 × RC-19; days to initiation of flowering (8.58%, 6.52%), days to physiological maturity (7.02%, 2.23%) and test weight (7.94%, 5.59%) in F<sub>2:3</sub> population of KBC-9  $\times$  PGCP-6. Similar findings were also recorded by Sabale et al. (2018) and Dinesh et al. (2017a). This indicates the presence of low genetic variability and non additive gene action for these traits. Hence, there is a limited scope for selection for traits with low variability. i.e., selection is not effective for these traits.

High heritability and GAM were recorded for plant height (94.41%, 40.96%), number of primary branches/plant (92.73%, 54.02%), number of pods/plant (83.79%, 31.99%), pod length (80.31%, 23.76%), number of seeds/ pod (84.38%, 27.95%), Hussainbi et al., Biological Forum – An International Journal 15(10): 580-584(2023)

dry matter/plant (95.88%, 44.35%) and seed yield/plant (94.14%, 53.29%) in  $F_{2:3}$  population of VBN-1  $\times$  RC-19; plant height (86.60%, 32.32%), number of primary branches/plant (95.58%, 62.79%), number of pods/plant (95.79%, 53.89%), number of seeds/ pod (82.91%, 24.64%), dry matter/plant (91.16%, 46.06%) and seed yield/plant (97.52%, 63.82%). Similar findings were reported by Verma et al. (2019), Meenatchi et al. (2019), Khan et al. (2013), Nair et al. (2018), Sheela (2017), Sabale et al. (2018), Mahesh et al. (2017), Bhadru and Navale (2012), Dinesh et al. (2017a), Dinesh et al. (2017b) and Khan et al. (2015). This indicates that the character is least influenced by the environment due to the presence of additive gene action. Here, selection based on phenotypic values for these traits could be more reliable.

Moderate heritability and GAM were recorded for days to initiation of flowering (47.48%, 10.58%) in  $F_{2:3}$ population of VBN-1 × RC-19; days to initiation of flowering (57.73%, 10.20%) in  $F_{2:3}$  population of KBC-9 × PGCP-6 indicating the presence of additive and non-additive gene action. Similar findings were recorded by Nair *et al.* (2018) and Verma *et al.* (2019). High heritability and moderate GAM were reported for *ournal* **15(10): 580-584(2023) 582**  days to physiological maturity (68.54%, 10.48%) and test weight (62.5%, 16.58%) in F<sub>2:3</sub> population of VBN- $1 \times \text{RC-19}$  indicating the influence of additive and nonadditive gene action and less influence of the environment for expression of the trait. Low heritability and GAM were reported for days to physiological maturity (10.10%, 1.46%) in F<sub>2:3</sub> population of KBC-9  $\times$  PGCP-6 which indicates high environmental influence on this trait. Hence, selection will be difficult due to the masking effects of the environment on genotypic effects.

Estimates of PCV, GCV, heritability and genetic advance as per mean were moderate to high for days to physiological maturity, plant height, number of primary branches per plant, number of pods per plant, number of seeds per pod, pod length, test weight, dry matter per plant and seed yield per plant for both the F<sub>2:3</sub> populations except days to initiation of flowering and days to physiological maturity in  $F_{2:3}$  populations of VBN-1  $\times$  RC-19; days to initiation of flowering, days to physiological maturity and test weight in F2:3 populations of KBC-9  $\times$  PGCP-6, this indicating that these traits are predominantly governed by additive genetic variance and parents are diverse with respect to most of the traits studied. Hence, there is a scope for selection for these traits with moderate to high variability. *i.e.*, selection is effective for these traits.

Moderate PCV and low GCV were observed for days to initiation of flowering in  $F_{2:3}$  populations of VBN-1 × RC-19. While, Low PCV and GCV were observed for days to physiological maturity in  $F_{2:3}$  populations of VBN-1 × RC-19; days to initiation of flowering, days to physiological maturity and test weight in  $F_{2:3}$ populations of KBC-9 × PGCP-6, indicating the presence of low genetic variability and non additive gene action for these traits. Hence, there is a limited scope for selection for traits with low variability. *i.e.*, selection is not effective for these traits.

## CONCLUSION

High phenotypic coefficient of variance and genetic coefficient of variance were recorded for plant height, number of primary branches/plant, dry matter/plant and seed yield/plant in  $F_{2:3}$  population of VBN-1 × RC-19. High heritability coupled with medium genetic advance as per cent of mean indicated the presence of both additive and non-additive gene actions for the inheritance of these characters and high genotype × environment interaction.

## FUTURE SCOPE

These characters could be improved suitably by modified selection procedure for mixed effects of additive and non additive gene actions like cyclic hybridization followed by selection. These characters and simple selection helps in development of high yielding cowpea genotypes.

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#### Conflict of Interest. None.

#### REFERENCES

- Bhardu, D. and Navale, P. A. (2012). Genetic variability parameters in F<sub>2</sub> and F<sub>3</sub> populations of cowpea (*Vigna unguiculata* L. Walp.). Legume Res., 35(1), 75-77.
- Boukar, O., Fatokun, C. A., Roberts, P. A., Abberton, M., Huynh, B. L., Close, T. J. and Ehlers, J. D. (2015). Cowpea, In: De Ron AM (ed) Grain legumes, Hand Book of Plant Breeding. *Springer, New York*, pp. 219–250.
- Carvalho, M., Lino Neto, T., Rosa, E. and Carnide, V. (2017). Cowpea: a legume crop for a challenging environment. J. Sci. Food. Agric., 97, 4273-4284.
- Dinesh, H. B., Viswanatha, K. P., Lohithaswa, H. C., Pavan, R. and Singh, P. (2017a). Variability, correlation and path analysis studies in F<sub>3</sub> generation of cowpea [Vigna unguiculata (L.) Walp]. Int. J. Curr. Microbiol. Appl. Sci., 6(9), 1420-1428.
- Dinesh, H. B., Viswanatha, K. P., Lohithaswa, H. C., Pavan, R., Singh, P. (2017b). Genetic variability, correlation and path analysis studies in an early segregating generation of cowpea (Vigna unguiculata L. Walp). Int. J. Pure App. Biosci., 5(5), 1389-1395.
- Goncalves, A., Goufo, P., Barros, A., Dominguez-Perles, R., Trindade, H., Rosa, E. A. and Rodrigues, M. (2016). Cowpea (Vigna unguiculata L. Walp.), a renewed multipurpose crop for a more sustainable Agri food system: nutritional advantages and constraints. J. Sci. Food Agric., 96, 2941-2951.
- Johnson, H. W., Robinson, H. F. and Comstock, R. E. (1955). Estimates of genetic and environmental variability in soybean. Agron. J., 47, 314-318.
- Khan, H., Vishwanatha, K. P. and Sowmya, H. C. (2013). Genetic variability and association studies in cowpea (*Vigna unguiculata* L. walp.). J. Food Legumes, 26(3 & 4), 42-45.
- Khan, H., Viswanatha, K. P. and Sowmya, H. C. (2015). Study of genetic variability parameters in cowpea (*Vigna unguiculata* L. Walp.) germplasm lines. *The Bioscan*, 10(2), 747-750.
- Mahesh, S., Sharma, P. P., Hemlata, S. and Deva, R. M. (2017). Genetic variability in cowpea (Vigna unguiculata L. Walp.) germplasm lines. J. Pharmacogn. Phytochem., 6(4), 1384-1387.
- Meenatchi, T., Thangaraj, K., Gnanamalar, R. P. and Pushpam, K. (2019). Genetic variability and heritability study on yield and its component traits in segregating population of cowpea (Vigna unguiculata L. Walp.). Electron. J. Pl. Breed., 10(2), 736-741.
- Munoz, A. M., Mirebrahim, H., Xu, P., Wanamaker, S. I., Luo, M., Alhakami, H. and Bozdag, S. (2017). Genome resources for climate resilient cowpea, an essential crop for food security. *Pl. J.*, 89, 1042– 1054.
- Nair, K. R., Desai, S. S., Sawardekar, S. V. and Burondkar, M. M. (2018). Study of Genetic Variability Parameters in F<sub>2</sub> Generation of Interspecific Hybrids in cowpea. *Int. J. Pure App. Biosci.*, 6(1), 954-958.
- Sabale, G. R., Bhave, S. G., Desai, S. S., Dalvi, M. B., Pawar, P. R. (2018). Variability, heritability and genetic advance studies in cowpea (Vigna unguiculata L.). Int. J. Curr. Microbiol. Appl. Sci., 7(9), 3314-3320.

- Sebetha, E. T., Modi, A. T. and Owoeye, L. G. (2014). Cowpea crude protein as affected by cropping system, site and nitrogen fertilization. *J. Agric. Sci.*, *7*, 224.
- Shanko, D., Andargie, M. and Zelleke, H. (2014). Interrelationship and path coefficient analysis of some growth and yield characteristics in cowpea (*Vigna unguiculata* L. Walp.) genotypes. J. Pl. Sci., 2(2), 97-101.
- Sheela, H. (2017). Variability studies in segregating populations of cowpea (Vigna unguiculata L. Walp). Ph. D. Thesis, Univ. Agric. Hortic. Sci., Shivamogga, India.
- Timko, M. P. and Singh, B. B. (2008). Cowpea, a multifunctional legume. *Genomics of Tropical Crop Plants, Springer*, New York pp. 227-258.
- Verma, A. K., Mehta, A. K., Gontia, A. S., Sharma, D., Singh, R. P. and Singh, P. (2019). Genetic variability, heritability and genetic advance studies for yield components in F<sub>2</sub> generation of cowpea (*Vigna unguiculata* L. Walp.). Int. J. Chem. Stud., 7(6), 3084-3088.

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