

## Algal Meal's Impact on the Fertilisation Abilities of Common Carp (*Cyprinus carpio* var. *communis*)

Hafsa Javeed<sup>1\*</sup>, Farooz Ahmad Bhat<sup>1</sup> and Oyas Asimi<sup>2</sup>

<sup>1</sup>Division of Fisheries Resource Management,

Sher-e- Kashmir University of Agricultural Sciences and Technology of Kashmir (J&K), India.

<sup>2</sup>Division of Fish Nutrition and Biochemistry, Sher-e- Kashmir University of Agricultural Sciences and Technology of Kashmir (J&K), India.

(Corresponding author: Hafsa Javeed\*)

(Received: 10 January 2023; Revised: 12 February 2023; Accepted: 20 February 2023; Published: 22 March 2023)

(Published by Research Trend)

**ABSTRACT:** *Spirulina* is a blue-green algae that contains upto 70% dry weight protein and is a good source of vitamins and PUFA. This study's goal was to evaluate the reproductive efficiency of Common carp fed with *Spirulina* supplemented diets at varied levels and to achieve the most effective supplementation of *S. platensis* in feed. The experiment was set up in which 150 *C. carpio* fish were assigned randomly to follow treatments: T<sub>1</sub> = *Spirulina* 25g/kg, T<sub>2</sub> = *Spirulina* 50g/kg, T<sub>3</sub> = *Spirulina* 75g/kg, T<sub>4</sub> = *Spirulina* 100g/kg and T<sub>0</sub> = feed without supplementation as control in 3 replicates. The results showed that supplementation of *Spirulina platensis* affected reproductive parameters in common carp and better results were obtained at a dosage of 7.5% i.e. T<sub>3</sub>.

**Keywords:** Algae, Fecundity, Feed, Gonadosomatic index, *Spirulina*.

### INTRODUCTION

Fish is a low-cost and high-quality source of lean meat. One of the most critical elements determining cultured fish's capacity to express their genetic capacity for development and reproduction is nutrition (Kumar and Ram 2016). Successful reproduction needs sufficient resources to support the high energy requirements for gamete development and reproduction (Volkoff and London 2018). The sum of the body's energy reserves influences the onset of reproduction and this reproductive process is susceptible to various metabolic factors. Gonadotropin-releasing hormone expression and release are regulated by metabolic hormones and neuropeptides that act on the hypothalamus centre represent the neuroendocrine system underlying the connection with relation to fertility and energy balance. Therefore, for the hypothalamus-pituitary-gonadal axis to fully activate throughout puberty and to function normally in maturity, sufficient bodily energy stores are required. In teleosts and other vertebrates, the hypothalamus-pituitary-gonadal axis regulates reproduction. Gonadotropin-releasing hormone, which controls pituitary gonadotropin production and release, as well as follicle-stimulating hormone and luteinizing hormone, are all produced by the pituitary. Gonadotropins act on the gonads to promote gonadal development through the release of sex steroid hormones. The brain and pituitary receive input from these steroids, completing the hypothalamic pituitary axis and regulating the reproductive cycle (Yaron *et al.*, 2003; Zohar *et al.*, 2010). As a result, the hypothalamic

gonadotropin-releasing hormone is thought to be a key player in teleost reproduction control.

In general, a large amount of food favours reproduction, while a small amount of food prevents the efficient functioning of the reproductive system (Schneider, 2004). The physiological mechanisms that split energy into different tasks during energetic difficulties favour processes that aid in survival over processes that aid in the development, longevity and reproduction (Shahjahan *et al.*, 2014). Proteins, carbohydrates and lipids are the three different food classes that the body uses to provide energy for various physiological processes and physical activities. The ability of different fish species to use energy-producing nutrients varies greatly. This difference is due to their normal feeding habits, which can be categorized as herbivorous, omnivorous and carnivorous. There is a link between natural eating habits and the need for protein in the diet as a result. Compared to herbivorous and omnivorous species, some carnivorous species need more protein in their diet. Carnivorous species excel at converting dietary fat and protein into energy, while they struggle with doing the same with carbohydrates.

### MATERIAL AND METHODS

#### A. Biochemical analysis of aquatic weeds

The Association of Official Analytical Chemists' methodologies were used to analyse the aquatic weeds for the proximate composition of several nutrients. Each determination was made three times. Fish meal was substituted with spirulina due to its high protein content AOAC (1990).

### B. Diet formation and preparation (Elghany, 2003)

Five experimental diets (control and four treatment groups) with 30% crude protein were formulated. Each diet's dry ingredients were well combined and dough was prepared by mixing the mixture with distilled water (100ml/kg). The ingredients were thoroughly blended using kitchen blender to make a paste of each diet. Four diets with different Spirulina concentrations were prepared (Table 1). The diet control (C) had no Spirulina,

similarly four treatment diets designated as T<sub>1</sub>, T<sub>2</sub>, T<sub>3</sub> and T<sub>4</sub> were formulated containing Spirulina at the rate of 2.5%, 5% and 7.5% and 10% respectively in addition to the basal feed ingredients. Pelleting of diets was carried out by passing the blended mixture through hand pelletizer with 1mm diameter mesh. All the five diets were dried and stored separately in air-tight containers for further use as described by Elghany (2003).

**Table 1: Percentage composition of experimental diets.**

Ingredient	Inclusion rate (%)					
	Protein (%)	Control (C)	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>
Fish meal (gm)	60	29	26.5	24	21.5	19
Mustard oil cake (gm)	45	22	22	22	22	22
Rice bran (gm)	13	25	25	25	25	25
Wheat flour (gm)	15	11	11	11	11	11
Vegetable oil (gm)	-	8	8	8	8	8
Vitamin & Mineral mixture (gm)	-	5	5	5	5	5
Spirulina (gm)	70	0	2.5	5	7.5	10
Total (gm)		100	100	100	100	100
<b>Chemical composition</b>						
Dry matter	-	89.5	90.4	90.5	90.6	89.5
Crude protein	-	30.3	30.7	30.8	30.9	30.3
Ether extract	-	8.05	7.71	7.45	7.11	8.05
Crude fiber	-	2.57	2.42	2.26	2.34	2.57
Ash	-	6.01	5.91	5.56	5.71	6.01

### C. Experimental Site

The experiment was performed in the Faculty of Fisheries' Wet Laboratory, Rangil Gandersbal, India, during the course of 90 days from April 1 to June 1 of this year. The fish were raised in plastic containers after a 10-day acclimation period.

### D. Experimental set-up and experimental fish

The experiment was carried out in the Fisheries farm Shuhama, India for 90 days 15 plastic circular tubs with a 80L capacity, covered with net. Initially, the tubs were cleaned and filled with a 4mgL<sup>-1</sup> solution of potassium permanganate. The following day, fresh water was used to properly wash the tubs and flush them out. Common carp, *Cyprinus carpio* specimens with average weight of 50g procured from fish farm shuhama were utilised in the investigation.

150 fish were randomly assigned to each of the four different experimental groups. Three replicates were used in each group, according to a totally random design. In each plastic tub, 80L of chlorine-free canal water was filled with ten fish, with an initial average weight of 50±5 g. There was constant aeration available. To evaluate the growth, the body weight was taken at intervals of 15 days. Before weighing the fish, they were starved overnight.

### E. Feeding

For the duration of the 60-day feeding trial, feeding was done at 5% of body weight. The daily diet was divided into two equal portions and given between 10:00 am and 6:00 pm.

### F. Reproductive biology

**Gonado-Somatic Index (GSI)** (Desai, 1970)

In order to study the gonad somatic index, the weight of the each fish specimen and corresponding ovary were calculated to 0.001 g precision. The ratio was computed through formula given by (Desai, 1970).

$$\text{Gonado Somatic Index} = \frac{\text{Weight of gonads (g)}}{\text{Total weight of fish (g)}} \times 100$$

### G. Fecundity Absolute fecundity

$$\text{Absolute Fecundity} = \frac{\text{No. of ova in the subsample} \times \text{Total Ovary weight}}{\text{Weight of subsample}}$$

For the estimate of absolute and relative fecundity, sub samples were obtained from the front, middle, and posterior areas of the ovary.

### H. Relative fecundity

$$\text{Relative Fecundity} = \frac{\text{Absolute Fecundity}}{\text{Weight of fish}}$$

### I. Statistical analysis

The data generated was analysed by appropriate statistical methods of Microsoft Excel and PAST for Windows (Version 20).

## RESULTS AND DISCUSSION

In males, the highest mean ± S.D. value of 5.03±1.22 for gonadosomatic index (GSI) was recorded in treatment group (T<sub>3</sub>) and the lowest mean ± S.D value of 3.88±1.9 for gonadosomatic index (GSI) was recorded in control group (T<sub>0</sub>). No significant difference (p>0.05) was seen in gonadosomatic index (GSI) among various treatment groups.

In females the highest mean ± S.D value for gonado somatic index (GSI) recorded in treatment group T<sub>3</sub> (9.43±3.86) and the lowest mean ± S.D value for gonado

somatic index (GSI) recorded in control group T<sub>0</sub> (3.71±2.13). Significant difference (p<0.05) was seen in gonado somatic index (GSI) of females among various treatment groups.

The gonadosomatic index (GSI) is a key indication of an organism's overall health. It's a tool for calculating or measuring reproductive capacity. Reproduction is the most important step in a species life cycle as it affects the species survival and success. The gonado somatic index (GSI) is a good predictor of changes in a fish's nutritional and energetic status (Adams *et al.*, 1996). The proportion of a fish's gonad weight to body weight is known as the gonad somatic index (GSI). As the ovaries of gravid females rapidly grow in size shortly prior to spawning, the gonad somatic index (GSI) is particularly useful in determining days and seasons of spawning. During the present study, the highest mean gonad somatic index (GSI) for both male (5.03) and female (9.43) was recorded in treatment group (T<sub>3</sub>) and the lowest mean gonado somatic index (GSI) value for both male (3.88) and female (3.71) was recorded in control group (T<sub>0</sub>). The fact that *Spirulina* contains sizable amounts of protein, lipids, and fatty acids—the primary components of egg yolk—could account for the treatment group T<sub>3</sub>'s highest mean gonado somatic index (GSI) value. Additionally, the necessary fatty acids in it give spawning energy. Dahlgren (1980) showed that the lipids, essential fatty acids, and proteins included in fish diets affect the rate at which vitellogenesis develops and oocytes mature. The improvement in the reproductive efficiency of treatment group (T<sub>3</sub>) may also be due to the fact that *Spirulina* has a good content of micronutrients (B group vitamins), which is in agreement with the findings of Coves *et al.* (1990) who explained the importance of B vitamins in elevating reproductive performance of fish. James *et al.* (2004) reported that female *Xiphophorus helleri* fed with 45% protein diets (animal and plant protein diets) had a greater ovary weight and gonado somatic index than those fed low protein diets (10-35%). Khanzadeh *et al.* (2016) also reported that the fish fed *Spirulina platensis* meal had greater gonad somatic indices than control. The outcomes of the current investigation concur with those of Guroy *et al.* (2012). *Spirulina* increased seed production in the golden tail cichlid (*Pseudotropheus acei*), gold fish (*Carrasius auratus*), swordtail (*Xiphophorus helleri*), and bassa fish (*Pangasius bocourti*), according to Vasudevan *et al.* (2011); James *et al.* (2006).

#### A. Absolute Fecundity

The highest mean±S.D. value of 5888±643.7 for absolute fecundity was observed in treatment group (T<sub>4</sub>) and the lowest mean±S.D. value of 3632±749.3 for absolute fecundity was observed in control group (T<sub>0</sub>). Statistically it was seen that there was significant difference (p<0.05) between the absolute fecundity of various treatment groups.

#### B. Relative fecundity

The highest mean±S.D. value of 99±13.37 for relative fecundity was observed in treatment group T<sub>3</sub> and the lowest mean±S.D. value of 69±13.08 for relative

fecundity was observed in control group T<sub>0</sub>. Significant difference (p<0.05) was seen in relative fecundity between various treatments groups.

The estimated number of eggs that will be laid during a spawning season is referred to as a fish's fecundity (Bagenal, 1957). Fecundity or reproductive potential is an important biological metric to consider when assessing the commercial potential of fish stocks (Gomez *et al.*, 2003). To understand the ability of fish populations to recover, accurate fecundity evaluation is necessary for effective fisheries management, including practical aquaculture (Tracey *et al.*, 2007). Fecundity and its relationship to female size makes it feasible to estimate egg output potential (Chondar, 1977), as well as the number of off springs that could be produced in a season and the reproductive capability of fish populations (Qasim and Qayyum 1963). In the study of fish biology and population dynamics, descriptions of reproductive techniques and assessments of fecundity are essential issues (Hunter *et al.*, 1992). Studies on reproduction, such as size at maturity, fecundity, reproductive season length, spawning behaviour and spawning fraction allow for the estimation of a fish's reproductive capacity. This data combined with estimations of egg production, allows for the calculation of spawning stock biomass (Saville, 1964; Parker, 1980; Laqsker, 1985). Even within a stock, fecundity is known to vary periodically, to change over time and to be proportional to fish size and hence age and condition (Kjesbu *et al.*, 1998). In both absolute and relative terms to body mass, larger fish produce more eggs. Fish size and condition are thus critical factors in determining population fecundity.

The absolute fecundity of Common carp in the present study ranged from 5888±643.7 to 3632±749.3. The relative fecundity varied from 99±13.37 to 69±13.08. To promote sexual development, reproduction, high-quality eggs, and larvae survival rates, the meals' essential fatty acids, alpha-tocopherol, ascorbic acid, and carotenoids were successful (Scabini *et al.*, 2011). James *et al.* (2006) found comparable outcomes in swordtail fish fed 8% spirulina. According to Bustamante and Ortega-salas (2002), the fecundity rate for three spotted gourami fish ranged from 8021 to 9104 eggs (2002). Furthermore, according to Guroy *et al.* (2012), feeding cichlid yellow tail (*Pseudotropheus acei*) 2.5% spirulina resulted in the largest number of eggs laid and hatching rates, which was a significant difference from other treatments.

## CONCLUSION AND FEATURE SCOPE

Gonado somatic index (GSI) of treatment group (T<sub>3</sub> @7.5% *Spirulina*) performed better than control (T<sub>0</sub>) for both males and females. Fecundity was recorded in terms of absolute and relative fecundity. The absolute fecundity of common carp in the present study ranged from 5888±643.7 to 3632±749.3 and the relative fecundity varied from 99±13.37 to 69±13.08. *Spirulina platensis* has beneficial effects on the growth, reproduction and blood physiology of common carp (*Cyprinus carpio* var. *communis*) based on the results of the present study. *Spirulina* can replace upto 7.5% (T<sub>3</sub>) of fish meal in the diet of common carp without any

adverse effects on growth. Currently, Spirulina production is directed towards its use as a feed additive at low inclusion levels. However, the present study demonstrates the feasibility of its use as a substitute source of protein in fish diets, giving arguments for intensification in Spirulina cultivation and its use in fish feeds.

**Conflict of Interest.** None.

## REFERENCES

- Adams, S.M., Ham, K.D., Greeley, M.S., LeHew, R.F., Hinton, D. E. and Saylor, C. F. (1996). Downstream gradients in bioindicator responses: point source contaminant effects on fish health. *Canadian Journal of Fisheries and Aquatic Sciences*, 53(10), 2177-2187.
- AOAC (1990). Official Method of Analysis of the Association of Official Analytical Chemists, 15<sup>th</sup> edition. AOAC, Arlington, VA. pp. 2220.
- Bagenal, T. B. (1957). The breeding and fecundity of the long rough dab *Hippoglossoides platessoides* (Fabr.) and the associated cycle in condition. *Journal of the Marine Biological Association of the United Kingdom*, 36(2), 339-375.
- Bustamente, R. H. and Ortega-salas, A. A. (2002). Initial sexual maturity and fecundity of two anabantids under laboratory conditions. *North American journal of aquaculture*, 64(3), 224-227.
- Chondar, S. L. (1977). Fecundity and its role in racial studies of *Gudusia chapra* (Pisces: Clupeidae). *The Proceedings of the Indian Academy of Sciences*, 86, 245-254.
- Coves, D., Audineau, P. and Nicolas, J. L. (1990). Rotifer Rearing Technology. In: *Aquaculture, Volume 1, Barnabe, G. (Ed.) Ellis Harwood, West Sussex, UK*, 232-245.
- Dahlgren, B. T. (1980). The effects of three different dietary protein levels on the fecundity in the guppy, *Poecilia reticulata* (Peters). *Journal of Fish Biology*, 16(1), 83-97.
- Elghany, A. E. (2003). Replacement of herring fish meal by soybean flour in practical diets for red tilapia, *Oreochromis niloticus* x *O. mossambicus*, grown in concrete tanks. *J. Applied Aquacult*, 14(1-2), 69-87.
- Gomez-Márquez, J. L., Peña-Mendoza, B., Salgado-Ugarte I H. and Guzmán-Arroyo, M. (2003). Reproductive aspects of *Oreochromis niloticus* (Perciformes: Cichlidae) at Coatetelco lake, Morelos, Mexico. *Revista de Biología Tropical*, 51(1), 221-228.
- Guroy, B.S., Ahin, I., Mantoglu, S and Kayal, S. (2012). *Spirulina* as a natural carotenoid source on growth, pigmentation and reproductive performance of yellow tail cichlid *Pseudotropheus acei*. *Aquac. Int. Springer Sci. Bus*, 20, 869–878.
- Hunter, J. R., Maciewicz, B. J., Lo, N. C. H. and Kimbrell, C.A. (1992). Fecundity, spawning, and maturity of female Dover sole, *Microstomus pacificus*, with an evaluation of assumptions and precision. *Fisheries Bulletin U.S.*, 90, 101-128.
- James, R. and Sampath, K. (2004). Effect of animal and plant protein diets on growth and reproductive performance in an ornamental fish, *Xiphophorus helleri*. *Indian Journal of Fisheries*, 51(1), 75-86.
- James, R., Sampath, K., Thangarathinam, R. and Vasudevan, I. (2006). Effect of dietary *Spirulina* level on growth, fertility, coloration and leucocyte count in red Swordtail, *Xiphophorus helleri*. *The Israeli Journal of Aquaculture Bamidgah*, 58(2), 97-104.
- Khanzadeh, M., Fereidouni, A.E. and Berenjestanaki, S.S. (2016). Effects of partial replacement of fish meal with *Spirulina platensis* meal in practical diets on growth, survival, body composition, and reproductive performance of three-spot gourami (*Trichopodus trichopterus*) (Pallas, 1770). *Aquaculture international*, 24(1), 69-84.
- Kjesbu, O. S., Withames, P. R., Solemdal, P. and Walker, M. G. (1998). Temporal variations in the fecundity of Arcto-Norwegian cod (*Gadus morhua*) in response to natural changes in food and temperature. *Journal of Sea Research*, 40(3-4), 303-321.
- Kumar, M. and Ram, M. (2016). Toxicity of some heavy metals on blood characteristics of freshwater fish *Clarias batrachus*. *International Journal of Fisheries and Aquatic Studies*, 4(1), 85-89.
- Qasim, S. Z. and Qayyum, A. (1963). Fecundities of some freshwater fish. *Proceedings of the National Institute of Sciences of India*, 29, 373-382.
- Scabini, V., Fernández-Palacios, H., Robaina, L., Kalinowski, T. and Izquierdo, M. S. (2011). Reproductive performance of gilthead seabream (*Sparus aurata* L., 1758) fed two combined levels of carotenoids from paprika oleoresin and essential fatty acids. *Aquaculture Nutrition*, 17(3), 304-312.
- Schneider, J. E. (2004). Energy balance and reproduction. *Physiology & behavior*, 81(2), 289-317.
- Shahjahan, M., Kitahashi, T. and Parhar, I. S. (2014). Central pathways integrating metabolism and reproduction in teleosts. *Frontiers in endocrinology*, 5, 36.
- Tracey, S. R., Lyle, J. and Haddon, M. (2007). Reproductive biology and perrecruit analyses of striped trumpeter (*Latris lineata*) from Tasmania, Australia: Implications for management. *Fisheries Research*, 84(3), 358-368.
- Volkoff, H. and London, S. (2018). Nutrition and reproduction in fish. *Encyclopedia of reproduction*, 743-748.
- Yaron, Z. V. I., Gur, G. A. L., Melamed, P., Rosenfeld, H., Elizur, A. and Levavi-Sivan, B. (2003). Regulation of fish gonadotropins. *International review of cytology*, 225, 131-185.
- Zohar, Y., Muñoz-Cueto, J. A., Elizur, A. and Kah, O. (2010). Neuroendocrinology of reproduction in teleost fish. *General and comparative endocrinology*, 165(3), 438-455.

**How to cite this article:** Hafsa Javeed, Farooz Ahmad Bha and Oyas Asimi (2023). Algal Meal's Impact on the Fertilisation Abilities of Common Carp (*Cyprinus carpio* var. *communis*). *Biological Forum – An International Journal*, 15(3): 305-308.