

Biological Forum – An International Journal

15(2): 144-153(2023)

ISSN No. (Print): 0975-1130 ISSN No. (Online): 2249-3239

Analysis of Seed Yield and its Contributing Traits Inheritance in M₃ Generation of EMS Induced Black Gram (*Vigna mungo* L. Hepper) Mutants

Bhattu Rajesh Nayak¹, Raju Padiya², K. Srinivas Naik³ and G. Vijay Kumar¹* ¹Department of Genetics, Osmania University, Hyderabad (Telangana), India. ²Department of Biochemistry, Osmania University, Hyderabad (Telangana), India. ³Centre for Plant Molecular Biology, Osmania University, Hyderabad (Telangana), India.

(Corresponding author: G. Vijay Kumar*)

(Received: 24 December 2022; Revised: 26 January 2023; Accepted: 30 January 2023; Published: 07 February 2023)

(Published by Research Trend)

ABSTARCT: The present study was carried to analyse the EMS induced on seed yield and its contributing characters of blackgram (accession IC-436524) when raised during M₃ generation in Kharif season in the month of mid June-mid September, 2022 in the field located Kodad, Suryapet, Telangana. Demand for grain legumes has been increased recently due to their high protein and nutritional values. This resulted for the production of huge quantity of grains to meet the demand for increasing population like India. Genetic variability is prerequisite for isolating high yield blackgram genotypes. Mutation breeding is the simple, cheap and best practise to genetic variability in self pollinated crop like blackgram. In this study, EMS induced blackgram (accession IC-436524) mutants was used to identify the inheritance of high yielding mutants of M₂ progeny during M₃ generation along with Untreated seeds were used as (Control) and T-9 (as check, collected from ICAR-CRIDA, Hyderabad). Investigation revealed that top 5 mutants with high morphological variations in Plant height, Branches, leaves, Clusters, Pods, Seeds, Seed yield, root length and root nodules were observed. It clearly indicated that there are adequate amount of variability among the blackgram mutants due to EMS treatments.

Keywords: Blackgram, mutants, M₃ generation, Seed yield.

INTRODUCTION

Blackgram locally called as Urdbean, is an important legume crop (Family: Leguminacae) rich in proteins (about 26%). It is known as "Poor man's meat" due to its high nutritional value of proteins, minerals, vitamins etc. It is being used as dietary protein for most of the vegetarian population. Blackgram is an excellent alternative for high quality protein with good digetability in nature.

This is one of the important crops cultivated in South East Asia including India. Most favorable climate for its cultivation is semi arid region (Abraham et al., 2013) and used as mixed crop along with rice, wheat, maize etc. It helps in atmospheric nitrogen fixation through the soil. It is also used as fodder for animals. Among the pulses it has high culinary values and high content of carbohydrates, proteins, minerals, fats, vitamins, potassium, calcium, iron, amino acids. Optimum temperature required for black gram plant growth is 27°C-30°C. Mostly cultivated in Kharif season throughout India. Black gram requires relatively heavier soil than other pulse grams. It can be grown on a variety of soils ranging from sandy soils to heavy cotton soil. The most ideal soil for cultivation of black gram crop is well drained loamy soil with a pH of 6.5 to 7.8. Average seed yield of blackgram is 604 Kg/ha

which was low when compared to other pulse crops like field pea (912 Kg/ha, Singh *et al.* (2015), chick pea (889 Kg/ha, Raina *et al.*, 2019), lentil (705 Kg/ha, Laskar *et al.*, 2018) (Annual Report 2016–2017). In order to break the gap and to meet the growing demand for production of black gram, efforts are needed to develop the high yielding varieties through crop improvement programmes.

The scheme of crop improvement programme is to create variabilities in crops such as high yielding, disease resistant, drought resistant etc. As the blackgram is a self pollinated crop which is having lesser variability, it is very important to create variability (Arvind Kumar *et al.*, 2007; Ramya *et al.*, 2014).

Genetic variability

Genetic variability through mutations

Crop plants have been recognized for their human health benefits because of rich in bioactive compounds like phenols, sugars, oils, minerals, vitamins, flavonoids, carbohydrates, etc. (Zia-ul-Haq *et al.*, 2014; Unlukara, 2019). Mutations played very significant source in increasing the crop productivity and simultaneously enhanced world food security, as an effective tool in supplementing the existing germplasm for cultivar improvement *i.e.*, new food crop varieties embedded with various induced mutations have contributed to the significant increase of crop production (Vanniarajan *et al.*, 2017; Barshile, 2006; Singh and Singh 2001; Sanjay, 2012; Patil *et al.*, 2003). Mutagenic agents are used to induce mutations for creating variability particularly for isolating mutants with desirable characters of economic importance. Mutagens cause's genetic changes in an organism, break linkages and produces many new promising characters (Shah *et al.*, 2008; Usharani and Kumar 2015). Mutations provide an opportunity to create hitherto unknown alleles, so that the plant breeder does not remain handicapped due to limited allelic variation at one or more gene loci of interest (Goyal and Khan 2010).

Although earlier studies on mutagenesis in blackgram have been studied (Gautam et al., 1992; Sharma et al., 2005; Kouser et al., 2007; Kumar et al., 2019; Raina and Khan 2020; Raina et al., 2020); Wani (2017, 2021); Amin et al. (2019); Goyal et al. (2019), yet limited reports are available on induced viable morphological mutations. Morphological mutations play an important role to change the characteristics of any varieties, to build new ideotype for development of new varieties. Mutants produced through induced mutations when used in cross breeding programmes, found more productive in development of improved new varieties (Pawar et al., 2000). Induced mutations are extensively used to create genetic variability for developing new mutants with desirable agro-economical characters (Laskar et al., 2019).

In Mutation breeding programmes, choice of an effective and efficient mutagen will certainly increases the possibility of creating desired mutations. The lethal dose (LD50) utilized an assumption that lower doses of treatment which effect minimum impacts on the genome and rarely generates phenotypic changes; whereas high doses may produce multiple impacts on the genome which consistently produces aberrations or negative changes (Ariraman *et al.*, 2014). Among the chemical mutagens used for inducing mutational studies, Ethyl methane sulfonate has been found highly reactive in inducing mutations and efficient in creating diversity in agronomic traits of food crops including black gram (Gnanamurthy and Dhanavel 2014).

Major protein portion in blackgram is Lysine (Gill et al., 2017). Inheritance of quantitative characters is regulated by interaction of different genes with additive effects on phenotypic variabilities (Laskar et al., 2018). The FAO/IAEA Mutant Variety Database (https://mvd.iaea.org accessed on 24 January, 2021) records show that out of total 466 released mutant varieties of legumes, only 9 have been released in urdbean till date, indicating that the crop is less exploited for mutation breeding (Goyal et al., 2019). Hence, sustained efforts are required to create reproducible mutation protocols for the development of novel genes the regulates economically important traits (Laskar and Khan 2017).

So the present research study was performed to evaluate the EMS induced mutations on Seed yield and yield contributing traits like plant height per plant (cm), number of branches per plant, number of leaves per plant, number of clusters per plant, number of seeds per plant, Seed yield (g) per plant, 100 seed weight(g), root length(cm) and root nodules per plant characters were studied in M_3 generations.

MATERIALS AND METHODS

Collection of Seeds. Black gram accession IC-436524 were obtained from NBPGR regional centre Hyderabad and T9 (check) were obtained from ICAR-CRIDA.

A. Treatment of Seeds using EMS

 M_1 generation was raised by treating the seeds with EMS prepared at different known concentration *viz.*, 0.2%, 0.3%, 0.4%, 0.5% and Control (untreated EMS) along with T9(check). All agronomic practices were strictly followed during preparation of field, sowing and subsequent management of M_1 , M_2 and M_3 generations.

B. M1 generation

Data was prepared accordingly and mutants with high range of morphological characters for high yielding in each row of each concentration like plant height per plant (cm), number of branches per plant, number of leaves per plant, number of clusters per plant, number of seeds per plant, Seed yield(g) per plant, 100 seed weight(g), root length(cm) and root nodules per plant were screened and isolated in M_1 generation (raised by inducing EMS). Seeds of M_1 plants were harvested separately and preserved in paper bags for next generation studies (M_2 generation).

C. M_2 generation

Data was prepared accordingly and mutants with high range of morphological characters for high yielding like plant height per plant (cm), number of branches per plant, number of leaves per plant, number of clusters per plant, number of seeds per plant, Seed yield (g) per plant, 100 seed weight(g), root length(cm) and root nodules per plant in each row of each concentration were screened and isolated in M₂ generation (raised by M₁ progeny). Seeds of M₂ plants were harvested separately and preserved in paper bags for next generation studies (M₃ generation). Fifty seeds from each treatment viz., 0.2% mutants, 0.3% mutants, 0.4% mutants, 0.5% mutants, Control and T-9 (totally 300 seeds) were sown in field located at Kodad, Survapet, Telangana to evaluate the seed yield and yield contributing traits.

For raising M_2 generation total 180 seeds of 30 healthy seeds from each treatment were collected from high yielding mutant and control along with T9 and were sown with 10cm × 30 cm distance between plants per row in a field in RBD (Randomized Block Design) with three replications each along with control. Similar agronomical parameters as M_1 generation were practiced to raise M_2 generation. M_2 population was evaluated for agronomic and morphological characters by phenotypical observations which are yield and yield contributing traits Plants which have high quantitative characters with high yielding in each row of each concentration were separated and seeds from those plants were collected and data was prepared based on the yield produced and yield contributing characters. For raising M₃ generation total 300 seeds of 50 healthy seeds from each treatment were collected from high yielding mutant and control along with T9 and were sown with 10cm \times 30 cm distance between plants per row in a field in RBD (Randomized Block Design) with three replications each along with control. Similar agronomical parameters as M₁ generation were practiced to raise M₃ generation.

RESULTS AND DISCUSSION

The Analysis of variance (ANOVA) results in M_3 generation revealed that "Significant genotypic differences were found for all ten characters (plant height (cm), number of branches per plant, number of clusters per plant, number of pods per plant, number of seeds per pod, 100-seed" weight (g), seed yield (g/pl), root length (cm), and number of root nodules) evaluated (Table 1.), indicating that there is a great deal of genetic variability among the mutants evaluated. Indicating that the black gram mutant has sufficient diversity as a result of treatments.

 Table 1: The Effect of Ems Mutagenesis on the Yield And Yield-Contributing Features of the M3 Generation of Black Gram Mutants was Analyzed using Analysis of Variance (ANOVA).

						"]	ASSQ				
Source of	df	Characters									
Varaiation	ui	Pl.Ht(cm) No.Br/Pl No.L/Pl No.Cl/Pl No.F		No.P/Pl	No S/Pl	S.Y(g)/Pl	100 S.Wt(g)	R.L(cm)/Pl	No.R.N/Pl		
Replications	4	84.868	0.233	2.805	7.077	47.993	1541.945	2.312	0.314	18.493	932.209
treatments	5	265.668 **	0.784 **	21.481 **	26.742 **	437.704 **	6312.709 **	10.576 **	0.678 **	28.065 **	1500.80**
Mean		45.39	2.453	12.89	11.1	44.234	179.37	6.49	3.165	21.475	135.73
Error		33.813	0.102	3.57	1.93	43.95	473.599	0.549	0.088	4.674	224.835
SEd		3.678	0.202	1.195	0.879	4.193	13.764	0.469	0.187	1.367	9.483
CV%		12.81	13.03	14.66	12.52	14.99	12.13	11.41	9.36	10.07	11.05

*Significant at 0.05% and ** at 0.01 % level, respectively

 Table 2: Performance of Quantitative Characters of Blackgram mutants (0.2, 0.3, 0.4 and 0.5% EMS Concentration) in M3 generation.

0.2% Mutants	Pl.Ht(c m)	No. Br/Pl	No. L/Pl	No. Cl/Pl	No.P/Pl	No S/Pl	S.Y(g)/Pl	100 S.Wt(g)	R.L (cm)/Pl	No. R.N/Pl
I-IC-436524-M3	62.52	3.1	18.8	12.9	51.3	215.8	8.59655	3.988	26.35	170.6
VI-IC-436524-M3	61.17	2.9	17.1	9.90	38.2	213.5	8.132092	3.8019	25.10	176.0
V-IC-436524-M3	56.86	3.0	16.8	12.9	51.2	188.6	7.57849	3.6095	23.65	154.0
III-IC-436524-M ₃	45.75	2.3	13.0	9.9	34.0	168.6	6.44038	3.052	20.36	135.9
VIII-IC-436524-M ₃	44.24	2.3	13.8	11.3	44.5	146.2	5.75771	2.7615	18.13	115.9
"0.3% Mutants										
IX-IC-436524-M3	50.91	3.0	14.2	15.00	62	240.6	9.37003	3.5165	22.51	137.8
V-IC-436524-M3	50.38	2.6	13.1	16.7	67.1	242.9	9.066475	3.3595	22.68	135.7
II-IC-436524-M ₃	44.92	2.6	12.4	12.1	48.5	204.2	7.92666	3.025	20.57	126.2
I-IC-436524-M ₃	43.23	2.4	10.6	16.5	67.5	181.5	7.00685	2.703	20.32	133.3
III-IC-436524-M ₃	41.76	2.1	11.1	13.6	52.9	160.5	6.6287	2.867	20.16	134.9"
"0.4% Mutants										
III-IC-436524-M ₃	49.71	2.5	13.8	10.4	48.4	188.8	6.69358	3.5451	23.088	157.5
V-IC-436524-M3	45.2	2.4	12.3	10.0	38.5	174.2	6.176376	3.1947	21.841	137.1
IV-IC-436524-M3	48.47	2.4	12.7	10.2	39.1	161.3	5.767579	3.2127	21.231	136.8
VI-IC-436524-M ₃	41.88	2.1	10.5	8.9	46.625	145.2	4.9128	2.709	18.146	140.5
II-IC-436524-M ₃	28.56	1.5	7.7	5.6	23.4	102.2	3.517176	2.0658	14.574	88.7"
"0.5% Mutants										
IX-IC-436524-M3	35.05	2	9.9	9.7	33.2	135.1	4.45569	2.641	20.126	125.9
X-IC-436524-M ₃	32.75	1.9	10.6	8.3	30.3	129.4	4.36597	2.698	18.796	117.4
V-IC-436524-M3	33.71	2	11.1	7.4	31.3	123.5	4.22602	2.738	18.856	116.8
VII-IC-436524-M ₃	30.58	1.7	9.7	7.1	28.8	111.3	3.78668	2.381	16.139	103.4
VI-IC-436524-M ₃	30.74	1.8	9.5	8.6	32.2	104.7	3.61481	2.416	15.499	112
Control	35.49	1.90	10.32	9.44	36.46	130.10	4.35	2.76	18.73	120.29
T-9	36.46	1.88	10.28	8.82	37.70	128.02	4.39	2.67	17.72	122.04

Top Mutants	Pl.Ht(cm)	No.Br/Pl	No.L/Pl	No.Cl/Pl	No.P/Pl	No S/Pl	S.Y(g)/Pl	100 S.Wt(g)	R.L(cm)/Pl	No.R.N/Pl
0.2% I-IC-436524-M3	62.52	3.1	18.8	12.9	51.3	215.8	8.59655	3.988	26.35	170.6
0.3% IX-IC-436524-M ₃	50.91	3	14.2	15	62	240.6	9.37003	3.5165	22.51	137.8
III-IC-436524-M ₃	49.71	2.5	13.8	10.4	48.4	188.8	6.69358	3.445	23.088	157.5
0.5% V-IC-436524-M ₃	33.71	2	11.1	7.4	31.3	123.5	4.22602	2.738	18.856	116.8
Control	35.49	1.90	10.32	9.44	36.46	130.10	4.35	2.76	18.73	120.29
T-9	36.46	1.88	10.28	8.82	37.70	128.02	4.39	2.67	17.72	122.04

Table 3: Performance of top five Blackgram mutants based on seed yield and yield contributing traits.

Table 4: Mean performance and % increased/decreased over control (untreated) and T-9 for ten quantitative characters in 0.2, 0.3, 0.4 and 0.5% mutant, control (untreated), T-9 of Blackgram in M₃ generation.

Characters	0.2% Mutant	Control(untreated)	T-9	% increased/decreased over control(untreated)	% increased/decreased over T-9
Pl.Ht(cm)	62.52	35.49	36.46	76.16	71.48
No.Br/Pl	3.10	1.90	1.88	63.16	64.89
No.L/Pl	18.80	10.32	10.28	82.17	82.88
No.Cl/Pl	12.90	9.44	8.82	36.65	46.26
No.P/Pl	51.30	36.46	37.70	40.70	36.07
No S/Pl	215.80	130.10	128.02	65.87	68.57
S.Y(g)/Pl	8.60	4.35	4.39	97.62	95.82
100 S.Wt(g)	3.99	2.76	2.67	44.49	49.36
R.L(cm)/Pl	26.35	18.73	17.72	40.68	48.70
No.R.N/Pl	170.60	120.29	122.04	41.82	39.79

Characters	0.3% Mutant	Control(untreated)	T-9	% increased/decreased over control(untreated)	% increased/decreased over T-9
Pl.Ht(cm)	50.91	35.49	36.46	43.45	39.63
No.Br/Pl	3.00	1.90	1.88	57.89	59.57
No.L/Pl	14.20	10.32	10.28	37.60	38.13
No.Cl/Pl	15.00	9.44	8.82	58.90	70.07
No.P/Pl	62.00	36.46	37.70	70.05	64.46
No S/Pl	240.60	130.10	128.02	84.93	87.94
S.Y(g)/Pl	9.37	4.35	4.39	115.40	113.44
100 S.Wt(g)	3.52	2.76	2.67	27.41	31.70
R.L(cm)/Pl	22.51	18.73	17.72	20.18	27.03
No.R.N/Pl	137.80	120.29	122.04	14.56	12.91

Characters	0.4% Mutant	Control(untreated)	Т-9	% increased/decreased over control(untreated)	% increased/decreased over T-9
Pl.Ht(cm)	49.71	35.49	36.46	40.07	36.34
No.Br/Pl	2.50	1.90	1.88	31.58	32.98
No.L/Pl	13.80	10.32	10.28	33.72	34.24
No.Cl/Pl	10.40	9.44	8.82	10.17	17.91
No.P/Pl	48.40	36.46	37.70	32.75	28.38
No S/Pl	188.80	130.10	128.02	45.12	47.48
S.Y(g)/Pl	6.69	4.35	4.39	53.88	52.47
100 S.Wt(g)	3.45	2.76	2.67	24.82	29.03
R.L(cm)/Pl	23.09	18.73	17.72	23.27	30.29
No.R.N/Pl	157.50	120.29	122.04	30.93	29.06

Characters	0.5% Mutant	Control(untreated)	T-9	% increased/decreased over control(untreated)	% increased/decreased over T-9
Pl.Ht(cm)	33.71	35.49	36.46	-5.02	-7.54
No.Br/Pl	2.00	1.90	1.88	5.26	6.38
No.L/Pl	11.10	10.32	10.28	7.56	7.98
No.Cl/Pl	7.40	9.44	8.82	-21.61	-16.10
No.P/Pl	31.30	36.46	37.70	-14.15	-16.98
No S/Pl	123.50	130.10	128.02	-5.07	-3.53
S.Y(g)/Pl	4.23	4.35	4.39	-2.85	-3.74
100 S.Wt(g)	2.74	2.76	2.67	-0.80	2.55
R.L(cm)/Pl	18.86	18.73	17.72	0.67	6.41
No.R.N/Pl	116.80	120.29	122.04	-2.90	-4.29

PI.Ht(cm)-Plant Height, No.Br/PI-Branches Plant⁻¹, No.L/PI- Leaves Plant⁻¹, No.Cl/PI-Clusters Plant⁻¹, No S/PI- Number of seeds Plant⁻¹No.P/PI-Pods Plant⁻¹, S.Y(g)/PI-Seed Yield Plant⁻¹, 100 S.Wt(g)-100 Seed Weight, R.L(cm)/PI-Root length (cm) Plant⁻¹ and No.R.N/PI-Root Nodules Plant⁻¹⁰,

Fig.1. Blackgram mutants M3 generation field and traits observations



Fig. G. Observed seven pods per clusters/ plant in 0.2% Fig. H. Observed seven pods size in mutants Fig. I. Observed increased seeds per pods in mutant:

The character plant height (cm) recorded mean value in 0.2% mutant was 62.52, 35.49 in control (untreated) and 36.46 in T-9 (check). The % increase calculated for the character plant height recorded was 76.16 over control (untreated) and 71.48 over T-9 (check). The character number of branches per plant recorded mean value in 0.2% mutant was 3.10, 1.90 in control (untreated) and 1.88 in T-9 (check). The % increase calculated for the character number of branches per plant recorded was 63.16 over control (untreated) and 64.89 over T-9 (check). The character number of leaves per plant recorded mean value in 0.2% mutant was 18.80, 10.32 in control (untreated) and 10.28 in T-9 (check). The % increase calculated for the character number of leaves per plant recorded was 82.17 over control (untreated) and 82.88 over T-9 (check). The character number of clusters per plant recorded mean value in 0.2% mutant was 12.90, 9.44 in control (untreated) and 8.82 in T-9 (check). The % increase calculated for the character number of clusters per plant recorded was 36.65 over control (untreated) and 46.26 over T-9 (check). Significant increment of clusters per plant was also reported earlier by Kumar et al. (2009) in black gram; Hakande, (1992) in chickpea. The character number of pods per plant recorded mean value in 0.2% mutant was 51.30, 36.46 in control (untreated) and 37.70 in T-9 (check). The % increase calculated for the character number of pods per plant recorded was 40.70 over control (untreated) and 36.07 over T-9 (check). Our results improvement of pod number per plant were more at lower dose of EMS, meets with earlier reports by Ponaganti et al. (2022) in mustard; Raghavendra et al. (2021) in sorghum; Ajaz et al. (2008); Basu et al. (2008) in blackgram; Patil (2009) in cowpea; mung bean by Auti et al. (2007), and urd bean by Singh et al. (2000). The character number of seeds per plant recorded mean value in 0.2% mutant was 215.80, 130.10 in control (untreated) and 128.02 in T-9 (check). The % increase calculated for the character number of seeds per plant recorded was 65.87 over control (untreated) and 68.57 over T-9 (check). The character seed yield (g/pl) recorded mean value in 0.2% mutant was 8.60, 4.35 in control (untreated) and 4.39 in T-9 (check). The % increase calculated for the character Seed vield (g/pl) recorded was 97.62 over control (untreated) and 95.82 over T-9 (check). Similar yield improvement under induced mutagenic treatment in blackgram was also reported earlier by Tamilzhzrasi et al. (2021); Selvam and Elangaimannan (2010); Singh et al. (2016); Longnathan et al. (2000); Das and Chakraborty (1998); Mehandi et al. (2013); Chand (2001). The character 100 seed weight (g) recorded mean value in 0.2% mutant was 3.99, 2.76 in control (untreated) and 2.67 in T-9 (check). The % increase calculated for the character 100 Seed weight (g) recorded was 44.49 over control (untreated) and 49.36 over T-9 (check) (Sinha et al., 2018; Murugan and Nadarajan 2006; Thangavel and Thirugnanakumar 2011)". The character root length (cm) recorded mean value in 0.2% mutant was 26.35, 18.73 in control (untreated) and 17.72 in T-9 (check). The % increase calculated for the character root length (cm) recorded was 40.68 over control (untreated) and 48.70 over T-9 (check). The character number of root nodules per plant recorded mean value in 0.2% mutant was 170.60, 120.29 in control (untreated) and 122.04 in T-9 (check). The % increase calculated for the character number of root nodules per plant recorded was 41.82 over control (untreated) and 39.79 over T-9 (check). The character plant height recorded mean value in 0.3% mutant was 50.91, 35.49 in control (untreated) and 36.46 in T-9 (check). The % increase calculated for the character plant height (cm) recorded was 43.45 over control (untreated) and 39.63 over T-9 (check). The character number of branches per plant recorded mean value in 0.3% mutant was 3.00, 1.90 in control (untreated) and 1.88 in T-9 (check). The % increase calculated for the character number of branches per plant recorded was 57.89 over control (untreated) and 59.57 over T-9 (check). The character number of leaves per plant recorded mean value in 0.3% mutant was 14.20, 10.32 in control (untreated) and 10.28 in T-9 (check). The % increase calculated for the character number of leaves per plant recorded was 37.60 over control (untreated) and 38.13 over T-9 (check). The character number of clusters per plant recorded mean value in 0.3% mutant was 15.00, 9.44 in control (untreated) and 8.82 in T-9 (check). Our results meets with Tamilzhzrasi et al. (2021); Pathak et al. (2017); Gadakh et al. (2013); Khan et al. (2004). The % increase calculated for the character number of clusters per plant recorded was 58.90 over control (untreated) and 70.07 over T-9 (check). The character number of pods per plant recorded mean value in 0.3% mutant was 62.00, 36.46 in control (untreated) and 37.70 in T-9 (check). The % increase calculated for the character number of pods per plant recorded was 70.05 over control (untreated) and 64.46 over T-9 (check). Many authors have contributed to this field, including Ponaganti et al. (2022) in mustard; Raghavendra et al. (2021) in sorghum; Singh et al. (2006); Panigrahi et al. (2015); Wani. (2006) in blackgram. The character number of seeds per plant recorded means value in 0.3% mutant was 240.60, 130.10 in control (untreated) and 128.02 in T-9 (check). The % increase calculated for the character number of seeds per plant recorded was 84.93 over control (untreated) and 87.94 over T-9 (check). The character seed yield (g/pl) recorded mean value in 0.3% mutant was 9.37, 4.35 in control (untreated) and 4.39 in T-9 (check). The % increase calculated for the character seed yield (g/pl) recorded was 115.40 over control (untreated) and 113.44 over T-9 (check). Similar findings were reported earlier by Tamilzhzrasi et al. (2021); Bhattu et al. (2022); Wani et al. (2008); (Makeen et al., 2007; Sharma and Ahmed 1997). The character 100 seed weight (g) recorded mean value in 0.3% mutant was 3.52, 2.76 in control (untreated) and 2.67 in T-9 (check). The % increase calculated for the character 100 Seed weight (g) recorded was 27.41 over control (untreated) and 31.70 over T-9 (check). The character root length (cm) recorded mean value in 0.3% mutant was 22.51, 18.73 in control (untreated) and 17.72 in T-9 (check). The % increase calculated for the character root length (cm) recorded was 20.18 over control (untreated) and 27.03 over T-9 (check). The character number of root nodules per plant recorded mean value in 0.3% mutant was 137.80, 120.29 in control (untreated) and 122.04 in T-9 (check). The % increase calculated for the character number of root nodules per plant recorded mean value in 0.3% mutant was 137.80, 120.29 in control (untreated) and 122.04 in T-9 (check). The % increase calculated for the character number of root nodules per plant recorded was 14.56 over control (untreated) and 12.91 over T-9 (check).

The character plant height recorded mean value in 0.4% mutant was 49.71, 35.49 in control (untreated) and 36.46 in T-9 (check). The % increase calculated for the character plant height recorded was 40.07 over control (untreated) and 36.34 over T-9 (check). The character number of branches per plant recorded mean value in 0.4% mutant was 2.50, 1.90 in control (untreated) and 1.88 in T-9 (check). There was a 31.58-percent increase in the reported character-level number of branches per plant compared to the control group (untreated) and 32.98 over T-9 (check). The character number of leaves per plant recorded mean value in 0.4% mutant was 13.80, 10.32 in control (untreated) and 10.28 in T-9 (check). The estimated percentage increase in character leaves per plant was 33.72 over control (untreated) and 34.24 over T-9 (check). The character number of clusters per plant recorded means value in 0.4% mutant was 10.40, 9.44 in control (untreated) and 8.82 in T-9 (check). According to the research of Pathak et al. (2017); Khan et al. (2004); Gadakh et al. (2013) the trait clusters per plant was observed positive results at medium EMS doses. Number of clusters per plant reported a % increase of 10.17 from the control (untreated) and 17.91 from T-9. There were 48.40 pods per plant on average in the 0.4% mutant, 36.46 in the control (untreated), and 37.70 in the T-9 plant (check). Character number of pods per plant increased by 32.75 percent over control (untreated) and by 28.38 percent over T-9 (check). Mean values for character counts of seeds per plant were 188.80 in a 0.4% mutant, 130.10 in a control (untreated), and 128.02 in T-9 (check). The % increase calculated for the character number of seeds per plant recorded was 45.12 over control (untreated) and 47.48 over T-9 (check). The character seed yield (g/pl) recorded mean value in 0.4% mutant was 6.69, 4.35 in control (untreated) and 4.39 in T-9 (check). The % increase calculated for the character Seed vield (g/pl) recorded was 53.88 over control (untreated) and 52.47 over T-9 (check). Significant increment for the trait seed yield at higher mutagenic doses was observed earlier by Raghavendra et al. (2021) in sorghum; Parveen et al. (2012); Khattak et al. (2001); Gupta (2005); Suguna et al. (2017); Umaharan et al. (1997); Sinha et al. (2018); Manivannan (1999). The character 100 seed weight (g) recorded mean value in 0.4% mutant was 3.45, 2.76 in control (untreated) and 2.67 in T-9 (check). The % increase calculated for the character

Nayak et al.,

100 Seed weight (g) recorded was 24.82 over control (untreated) and 29.03 over T-9 (check). The character root length (cm) recorded mean value in 0.4% mutant was 23.09, 18.73 in control (untreated) and 17.72 in T-9 (check). The % increase calculated for the character root length (cm) recorded was 23.27 over control (untreated) and 30.29 over T-9 (check). The character number of root nodules per plant recorded mean value in 0.4% mutant was 157.50, 120.29 in control (untreated) and 122.04 in T-9 (check). The % increase calculated for the character number of root nodules per plant recorded mean value in 0.4% mutant was 30.93 over control (untreated) and 29.06 over T-9 (check).

The character plant height (cm) recorded mean value in 0.5% mutant was 33.71, 35.49 in control (untreated) and 36.46 in T-9 (check). The % decrease calculated for the character plant height recorded was -5.02 over control(untreated) and -7.54 over T-9 (check). The character number of branches per plant recorded mean value in 0.5% mutant was 2.00, 1.90 in control (untreated) and 1.88 in T-9 (check). Number of individual plant branches increased by 5.26 percent as compared to the control group (untreated) and 6.38 over T-9 (check). The character number of leaves per plant recorded means value in 0.5% mutant was 11.10, 10.32 in control (untreated) and 10.28 in T-9 (check). "The % increase calculated for the character number of leaves per plant recorded was 7.56 over control (untreated)" and 7.98 over T-9 (check). The character number of clusters per plant recorded mean value in 0.5% mutant was 7.40, 9.44 in control (untreated) and 8.82 in T-9 (check). Character number clusters per plant showed a % reduction of -21.61 when compared to the control(untreated) and -16.10 when compared to T-9 (check). The 0.5% mutant had a mean pod count of 31.30, 36.46 for controls and 37.70 for T-9 (check). The % decrease calculated for the character number of pods per plant recorded was -14.15 over control (untreated) and -16.98 over T-9 (check). The decreased number of pods per plant at higher doses of mutagen was observed earlier by Barshile, (2006) in chickpea and Sagade (2008) in urdbean. The character number of seeds per plant recorded means value in 0.5% mutant was 123.50, 130.10 in control (untreated) and 128.02 in T-9 (check). The % decrease calculated for the character number of seeds per plant recorded was -5.07 over control (untreated) and -3.53 over T-9 (check). The character seed yield (g/pl) recorded mean value in 0.5% mutant was 4.23, 4.35 in control (untreated) and 4.39 in T-9 (check). The % decrease calculated for the character seed yield (g/pl) recorded was -2.85 over control (untreated) and -3.74 over T-9 (check). The character 100 seed weight (g) recorded mean value in 0.5% mutant was 2.74, 2.76 in control (untreated) and 2.67 in T-9 (check). The % decrease calculated for the character 100 seed weight (g) recorded was -0.80 over control (untreated) and increased (2.55) over T-9 (check). The character root length (cm) recorded mean value in 0.5% mutant was 18.86, 18.73 in control (untreated) and 17.72 in T-9 (check). The % increase calculated for the character root length (cm) recorded was 0.67 over control (untreated) and 6.41 over T-9 (check). The character number of root nodules per plant recorded mean value in 0.5% mutant was 116.80, 120.29 in control (untreated) and 122.04 in T-9 (check). The % decrease calculated for the character number of root nodules per plant recorded was -2.90 over control (untreated) and -4.29 over T-9 (check). The reduction of different quantitative traits used higher dose of mutagens were reported earlier by Raghavendra et al. (2021) in sorghum; Pathak et al. (2017); Thomas and Sreekumar (2001); Singh et al. (1998); Aher et al. (2001); Suguna et al. (2017); Thangavel and Thirugnanakumar (2011); Srinives et al. (1991); Wani, (2006). The overall results revealed that top five mutants based on seed yield (0.2%) I-IC-436524-M3 (8.59g/pl), (0.3%) IX-IC-436524-M₃ (9.370g/pl), (0.4%) III-IC-436524-M₃ (6.693g/pl) and (0.5%) V-IC-436524-M₃ (4.45g/pl) in M₃ Generation were identified and also observed with six pods per cluster, big trifoliate green leaves, average plant height was observed was upto 70cm, 8-10 seeds per pod, black and healthy seeds are abundant per plant, more branches with tendril type and more clusters/plant have been observed in M₃ generation.

CONCLUSIONS

Among top five mutants, two mutants from 0.2% and 0.3% EMS *viz.*, I-IC-436524-M3 and IX-IC-436524-M3 showed highest seed yield (g/pl). The % increase calculated for the character seed yield (g/pl) of I-IC-436524-M3 (0.2%) recorded was 97.62 over control (untreated) and 95.82 % over T-9 (check). The % increase calculated for the character seed yield (g/pl) of IX-IC-436524-M3 (0.3%) recorded was 115.40% over control (untreated) and 113.44% over T-9 (check) in M3 generation. Hence these mutants could be utilized for further crop improvement programme in blackgram.

Acknowledgement. In the doctoral dissertation, the current research is included. We acknowledge Osmania University, Hyderabad for providing the lab and outdoor research locations. We also acknowledge the contribution of seed for the blackgram accession from ICAR-CRIDA, Hyderabad, and the NBPGR Regional Center, Hyderabad. Conflict of Interest. None.

REFERENCES

- Abraham, B. Vanaja, M. Reddy, P. R. Sunil, N. S. N. Kamala, V. and Vara Prasad, K. S. (2013). Identification of stable and high yielding genotypes in black gram [*Vigna mungo* (L.) Hepper] germplasm. *Indian Journal of Genetics*, *73*, 264–269.
- Aher, R. P. Dahat, D. V. and Surve, P. P. (2001). Diallel analysis for yield contributing characters in mungbean. *Legum Research*, 24(2), 124-126.
- Aijaz, A. Wani, and Mohammad, Anis. (2008). Gamma Rayand EMS-Induce Bold-seeded High- Yielding Mutants in Chickpea (*Cicer arietinum L.*), *Turkish Journal of Biology*, 32, 161-166.

- Amin Ruhul, Mohammad Rafiq Wani, Aamir Raina, Shahnawaz Khursheed and Samiullah Khan. (2019). Induced Morphological and Chromosomal Diversity in the Mutagenized Population of Black Cumin (*Nigella sativa* L.) Using Single and Combination Treatments of Gamma Rays and Ethyl Methane Sulfonate. Jordan Journal of Biological Sciences, 12(1): 23-30.
- Annual Report (2016–2017). In: Government of India, Ministry of Agriculture and Farmers Welfare, Department of Agriculture, Cooperation and Farmers Welfare, Directorate of Pulses Development, Vindhyachal Bhavan, India. Accessed 25 Aug 2018.
- Ariraman, M. Gnanamurthy, S. Dhanavelb, D. Bharathi, T. and Murugan, S. (2014). Mutagenic effect on seed germination, seedling growth and seedling survival of Pigeon pea (*Cajanus cajan* (L.) Millsp). *International Letters of Natural Science*, 16, 41-49.
- Arvind Kumar, M. N. Mishra and M.C. Kharkwal. (2007). Induced mutagenesis in blackgram [Vigna mungo (L.) Hepper]. Indian Journal of Genetics, 67(1), 41-46.
- Auti, S. G. Dalave, S. C. Patil, M. T. and B. J. Apparao. (2007). Effect of chemical and physical mutagens on Yield contributing characters in M₂ generation of Mungbean (*Vigna radiata* (L.) Wilzek): In Proceedings of Plant Science ences (Eds. Prof. S. M. Reddy, ISCA) Indian Science Congress (3-7 Jan (2007)) held at Annamalai University, Chidambaram (TN), India.
- Barshile, J. D. (2006). Studies on effect of mutagenesis employing EMS, SA and GR in Chickpea (*Cicer* arietinum L.). Ph.D. Thesis. University of Pune, Pune (MS), India. Unpubl.
- Basu, S. K. Acharya, S. H. and Thomas, E. J. (2008). Genetic improvement of fenugreek (*Trigonella foenum* graecum L.) through EMS induced mutation breeding for higher seed yield under western Canada prairie conditions. Euphytica, 160, 249-258.
- Bhattu Rajesh Nayak, K. Srinivas Naik, G. Kumara Joshi, G. Vijay Kumar (2022). Evaluation of Seed Yield and its Contributing Traits in Blackgram (Vigna mungo L. Hepper) Mutants Raised during M₂ Generation. International Journal of Food and Nutritional Sciences, 11(2), 1562-1573.
- Chand, P. (2001). Studies on gene action for yield and its components in urdbean [Vigna mungo (L) Hepper]. Legume Research-An International Journal, 24(2), 121-123.
- Das, S. V. and S. Chakraborty, (1998). Genetic variation for seed yield and its components in green gram (Vigna radiata (L.) Wilczek). Plant Science., 11, 271-273.
- Gadakh, S. S. Dethe, A. M. Kathale, M. N. and Kahate, N. S. (2013). Genetic diversity for field and its component traits in green gram [*Vigria radiate* (L.) Wilczek], 9, 106-109.
- Gautam, A. S. K. C. Sood and A. K. Ricarria, (1992). Mutagenic effectiveness and efficiency of gamma rays, ethylmethane sulphonate and their synergistic effect in blackgram (*Vigna mungo* L.). *Cytolog ia*, *5*, 85-89.
- Gill, R. K. Singh I. Kumar A and Singh S. (2017). Assessment of combining ability for various quantitative traits in summer Urd bean. *Electronic Journal of Plant Breeding*, 6, 412-416.

- Gnanamurthy, S. D. Dhanavel, T. Bharathi and N. Jagajanantham (2014). Effect of physical mutagens gamma radiation on the morphological characters of cowpea [Vigna unguiculata (L.)Walp]. International Journal of Current Research and Development, 1(1), 5-10.
- Goyal, S. Khan, S. (2010). Cytology of induced morphological mutants in *Vigna mungo* (L.) Hepper. *Egypt J Biol.*, 12, 81–85.
- Goyal, S. Wani, M. R. Laskar, R. A. Raina, A. Amin, R. & Khan, S. (2019). Induction of morphological mutations and mutant phenotypingin black gram (*Vigna mungo* (L.) Hepper) using gamma rays and EMS. *Vegetos*, 32, 464–472.
- Gupta, R. S. (2005). Genetic analysis for Yield and its component traits in Urd bean (*Vigna mungo* (L.) Hepper): Ph.D. Thesis, p 198, Narendra Deva University of Agriculture and Technology, Faizabad, India.
- Hakande, T. P. (1992). Cytological studies in *Psophocarpus* tetragonolobus (L.) PC. Ph.D. thesis, Marathwada University, Aurangabad, MS, India.
- Khan, S. Wani, M. R. Bhat, M. D. Parveen, K. (2004). Induction of mor-phological mutants in chickpea. *Int Chickpea Pigeonpea News-lett.*, 11, 6-7.
- Khattak, G. S. S. Haq, M. A. Ashraf, M. and Mc Nelly, T. (2001).Genetic basis of variation of Yield and its components in Mungbean (*Vigna radiata* (L.) Wilczek): *Heriditas*, 134, 211-217.
- Kouser, M. Babu, G. S. Lavanya, G. R. (2007). Effects of mutagen on M_1 population in urdbean. J Food Legum, 20(1), 109–110.
- Kumar, G. V. Vanaja, M. Lakshmi, N. J. and Maheswari, M. (2015). Studies on variability, heritability and genetic advance for quantitative traits in blackgram (*Vigna mungo* (L.) Hepper). *Agricultural Research Journal*, 52(4), 28-31.
- Kumar, V. Sharma, A.K. Singh, V.P. Kumar, M. (2009). Characterization of pre-breeding genetic stocks of urdbean (*Vigna mungo* (L.) Hepper) induced through mutagenesis. In: Shu QY (ed) Induced plant mutations in the genomics era. Food and Agri-culture Organization of the United Nation, Rome, pp 391– 394.
- Laskar, R. A. and Khan, S. (2017). Mutagenic Effectiveness and Efficiency of Gamma Rays and HZ with Phenotyping of Induced Mutations in Lentil Cultivars. *International Letters of Natural Sciences*, 64, 17-31.
- Laskar, R. A., Laskar, A. A., Raina, A., & Amin, R. (2018). Induced mutation analysis with biochemical and molecular characterisation of high yielding lentil mutant lines. *International Journal of Biological Macromolecules*, 109, 167–179.
- Laskar, R. A. Wani, M. R. Raina, A. Amin, R. and Khan, S. (2019). Morphological characterization of gamma rays induced multipodding (mp) mutant in lentil cultivar Pant L-406. *Int. J. Rad. Biol.*, 94, 1049–1053.
- Longnathan, P. Sarvan, K. and Ganesh, J. (2000). Genetic analysis of yield and related components in greengram (Vigna radiata L). Res. on Crop., 1(1), 34-36.

Nayak et al.,

- Makeen, K. Garad, A. Arif, J. and Archana, K.S. (2007). Genetic variability and correlation studies on Yield and its components in Mungbean (*Vigna radiata* (L.) Wilczek): *Journal of Agronomy*, 6, 216-218.
- Manivannan, N. (1999). Genetic variability for seed yield and its components of greengram (*Vigna radiata* (L.) Wilczek). Agric. Sci. Digest., 12(2), 96-90.
- Mehandi, S. Singh, C. M. and Kushwaha, V. L. (2013). Estimates of Genetic variability and Heritability for yield and yield contributing traits in Mung bean (*Vigna radiata* (L.) Wilczek). *The Bioscan*, 8, 1481-1484.
- Wani, M. R. (2006). Estimates of genetic variability in mutated populations and the scope of selection for yield attributes in *Vigna radiata* (L.) Wilczek. *Egyptian Journal of Biology*, 8.
- Murugan, E. and Nadarajan, N. (2006). Breeding for improved Yield and yellow mosaic virus disease resistance in Blackgram (*Vigna mungo* (L.) Hepper): Second National Plant Breeding Congress- Plant Breeding in Post Genomic Era, P 129-133.
- Panigrahi, K. K. Mohanty, A. Pradhan, J. Das, T. R. and Baisakh, B. (2015). Estimation of combining ability in Black gram (*Vigna mungo* (L.) Hepper) for yield and its attributing traits using diallel crossing method. *Electronic Journal of Plant Breeding*, 6, 631-657.
- Parveen, I. S. Redi, S. M. Reddy, D. M. and Suhakar, P. (2012). Heterosis and combining ability for Yield and Yield components in Urd bean (*Vigna mungo* (L.) Hepper): Journal of Agriculture and Allied Sciences, 3, 14-44.
- Pathak, A. R. Naik, M. R. and Joshi, H. K. (2017). Heterosis, inbreeding depression and Heritability for Yield and Yield components in cowpea. *Electronic Journal of Plant Breeding*, 8, 72-77.
- Patil, M. T. (2009). Genetic Improvement in cowpea (Vigna unguiculata (L.) for agronomic traits through mutation breeding. Ph. D. Thesis. University of Pune.
- Patil, S. Nair, B. Maheshwari, J. J. and Pillewan, S. (2003). Variability studies in M₂ and M₃ generation of soybean mutants. *Adv. Plant Sci.*, 16(1), 295-299.
- Pawar, S. E. Manjaya, J. G. Souframanien, J. Bhatia, C. R. (2000). Genetic improvement of pulse crops: induced mutations and their use in cross breeding. In: Proceedings FAO/IAEA workshop on the use of nuclear and molecular techniques in crop improvement. BARC, Mumbai, pp 170-174.
- Ponaganti Shiva Kishore, Aditya Pratap Singh, Sujaya Dewanjee and Pramod Kumar Pandey (2022). Mutation Breeding as a Tool for Aphid Resistance in Indian mustard [*Brassica juncea* (L.) Czern & Coss]. *Biological Forum – An International Journal*, 14(1), 1111-1118.
- Prasad, P. V. V. Djanaguiraman, M. Perumal, R. and Ciampitti, I. A. (2015). Impact of high temperature stress on floret fertility and individual grain weight of grain sorghum: sensitive stages and thresholds for temperature and duration. *Front. Plant Sci.*, 6, 820.
- Raghavendra, V. C. G. Girish, M. B. Ashok, B. V. Temburne, L. N. Yogeesh. (2021). Screening of M4 Sorghum (Sorghum bicolor (L.) Moench) Mutant Lines Against Shoot Fly (Atherigona soccata Rondani), Biological Forum – An International Journal, 13(3a): 774-780.

- Rai, K.N. Hanna, W.W. (1990). Morphological characteristics of tall and dwarf pearl millet isolines. *Crop Science*, 30, 23–25.
- Raina, A. Khan, S. Wani, M.R. Laskar, R.A. Mushtaq, W. (2019). Advances in Plant Breeding Strategies: Legumes. Springer; Cham: Chickpea (*Cicer* arietinum L.) cytogenetics, genetic diversity and breeding; 53–112.
- Raina, A. Khan, S. (2020). Rice Research for Quality Improvement: Genomics and Genetic Engineering. Springer; Singapore: Increasing rice grain vield under biotic stresses: mutagenesis, transgenics and genomics approaches; 149-178.
- Ramya, B. Nallathambi, G. (2014). Effect of mutagenesis on germina-tion, survival, pollen and seed sterility in M-1 generation of black gram (*Vigna mungo* (L) Hepper). *Plant Arch.*, 14(1), 499–501.
- Sagade, A. B. (2008). Genetic improvement of Urd bean through mutation breeding. Ph. D. thesis. University of Pune.
- Sanjay, G. and Auti, (2012). Induced morphological and quantitative mutagenesis in mungbean. Induced seed and pod colour mutations in urdbean (*Vigna mungo* (L.) Hepper). *Indian Journal of Genetics and Plant Breeding*, 67(3).
- Selvam, Y.A. and Elangaimannan, R. (2010). Combining ability analysis for yield and its component traits in Blackgram (Vigna Mungo (L.) Hepper): Electronic Journal of Plant Breeding, 1, 1386-1391.
- Shah, T. M. Mirza, J. I. Haq, M. A. Atta, B. M. (2008). Induced genetic variabil-ity in chickpea (*Cicer* arietinum L.). II. Comparative mutagenic effectiveness and efficiency of physical and chemical mutagens. *Pakistan Journal of Botany*, 40(2), 605– 613.
- Sharma, D. and Ahmed, N. U. (1997). Genetics and combining ability studies for Yield and its components in Black gram (Vigna mungo (L.) Hepper): Journal of Agricultural Science and Society North East India, 10, 19-24.
- Sharma, S. K. Sood, R. Pandey, D. P. (2005). Studies on mutagen sensitivity, effectiveness and efficiency in urdbean (*Vigna mungo* (L.) Hepper). *Indian Journal* of Genetics, 65, 20-22.
- Singh, A. K. and R. M. Singh (2001). Mutagenic efficiency and effectiveness of gamma rays, ethyl methane sulphonate and their combination in mung bean. *Crop. Improv.*, 28, 260-266.
- Singh, D. Gill, N.S. and Singh, K.B. (2015). Yield gap analysis of cotton crop through frontline demonstrations in central plain zone of Punjab. *Indian J. of Soc. Res.*, 56(2), 245-249.
- Singh, G. R. Sareen, P. K. Sareen and R. P. Saharan, (2000). Induced chlorophyll and morphological mutations in mungbean. *Indian Journal of Genetics*, 60(3), 391-393.
- Singh, I. S. Singh, B. D. Singh, R. P. and Singh, K. K. (1998). Interrelationship of yield and its components in F_3 progenies of a cross in mungbean. *Crop Improvement*, 15(2), 146-150.
- Singh, R. K. (1996). Gamma ray induced bold seeded mutant in Vigna mungo (L.) Hepper. Indian Journal of Genetics, 56(1), 104–108.

Nayak et al.,

- Singh, S. P. N. K. Singh, R. P. Singh and J. P. Prasad, (2006). Mutagenic effect of gamma rays and EMS on nodulation yield traits on Lentil. *Indian Journal of Pulses Research*, 19, 53-55.
- Singh, V. P. Singh, M. Pal, J. P. (1999). Mutagenic effects of gamma rays and EMS on frequency and spectrum of chlorophyll and macro-mutations in urdbean (*Vigna mungo* (L.) Hepper). *Indian Journal of Genetics*, 59(2), 203–210.
- Sinha, S. Mishra, S.B. Paramhans, P. and Pandey, S.S. (2018). Heterosis and inbreeding depression for Yield and Yield components in intraspecific crosses of Vigna. International Journal of Current Microbiology and Applied Sciences, 7, 4912-4917.
- Sonu Goyal and Samiullah Khan (2010). Induced Mutagenesis in Urdbean Vigna mungo L. Hepper): A Review, International Journal of Botany, 6(3), 194-206.
- Sonu Goyal, Mohammad Rafiq, Wani, Rafiul, Amin Laskar, Aamir Raina, Ruhul Amin, Samiullah Khan, (2019). Induction of morphological mutations and mutant phenotyping in black gram [Vigna mungo (L.) Hepper] using gamma rays and EMS. Vegetos, 32, 464–472.
- Srinives, P. Tangbunitivong, W. and Griffing, B. (1991). Genetic study of yield components in mungbean (*Vigna radiata* (L.) Wilczek) grown in dry and wet seasons, J. National Res. Council Thailand, 23(1), 1-13.
- Suguna, R. Savitha, P. and Ananda, C.R.K. (2017). Inheritance of Genetic variability, Combining Ability and Heterosis for Yellow Mosaic Virus Disease Resistance and Yield Improvement in Blackgram (Vigna mungo (L.) Hepper): International Journal of Current Microbiology and Applied Sciences, 6, 2416-2442.
- Tamilzhzrasi, M. Kumaresan, D. Souframanien, J. Jayamani, P. (2021). Effect of Induced Muatagenesis in M₁ generation of black gram (*Vigna mungo L. Hepper*). *Multilogic in Science: YC Journal.* Vol. X, Issue XXXIII.

- Thangavel, P. and Thirugnanakumar, S. (2011). Genetics of seeds Yield and its component characters in green gram (Vigna radiata (L.) Wilczek): Plant Archives, 11, 183-185.
- Thomas, B. and Sreekumar, S.G. (2001). Combining ability for biological nitrogen fixation traits and Yield components in Blackgram (*Vigna mungo* (L.) Hepper): *Journal of tropical agriculture*, 39, 93-97.
- Umaharan, P. Ariyanayagam, R.P. and Haque, S.Q. (1997). Genetic analysis of Yield and its components in vegetable cowpea (*Vigna unguiculata* L. Walp): *Euphytica*, 96, 207.
- Unlukara, A. (2019). Effects of depth-dependent irrigation regimes and organomineral fertilizers on water use and quality attributes of sugar beet. *Turk. J. Agric. For.*, 43, 492–499.
- Usharani, K.S. Kumar, C.R.A. (2015). Induced viable mutants in urdbean (*Vigna mungo* (L.) Hepper). *Bioscan*, 10(3), 1103–1108.
- Vanniarajan, C. S. Ganeshram, J. Souframanien, K. Veni, S. Anandhi Lavanya, and Kuralarasan, V. (2017). Gamma rays induced urdbbean [Vigna mungo (L.) Hepper] mutants with YMV resistance, good batter quality and bold seeded type. Legume res., DOI: 10.18805/LR-3824.
- Wani, M. R. (2017). Induced chlorophyll mutations, comparative mutagenic effectiveness and efficiency of chemical mutagens in lentils (*Lens culinaris* Medik). Asian J. Plant Sci., 16, 221–226.
- Wani, M.R. (2021). Comparative biological sensitivity and mutability of chemo-mutagens in lentil (*Lens culinaris* Medik), *Legume Res.*, 44, 26-30.
- Wani, M. R. Khan, S. (2014). Estimates of genetic variability in mutated populations and the scope of selection for yield attributes in *Vigna radiata* (L.) Wilczek. *Egypt. J. Biol.*, 2006; 8,1–6.
- Zia-ul-Haq, M. Ahmad, S. Bukhari, S. A. Amarowicz, R. Ercisli, S. Jaafar, H. Z. E. (2014). Compositional studies and biological activities of some mash bean (*Vigna mungo* (L.) Hepper) cultivars commonly consumed in Pakistan. *Biol. Res.*, 47, 23.

How to cite this article: Bhattu Rajesh Nayak, Raju Padiya, K. Srinivas Naik and G. Vijay Kumar (2023). Analysis of Seed Yield and its Contributing Traits Inheritance in M₃ Generation of EMS Induced Black Gram (*Vigna mungo* L. Hepper) Mutants. *Biological Forum – An International Journal*, *15*(2): 144-153.