

15(8): 561-566(2023)

ISSN No. (Print): 0975-1130 ISSN No. (Online): 2249-3239

Impact of Water Soluble Fertilizers and Silica on the Growth of African Marigold (*Tagetes erecta* L.)

Bharathkumar C.¹, A. Subbiah²*, D. Keisar Lourdusamy³, K. Sathiya Bama⁴ and K. Vanitha⁵¹Research Scholar, Department of Floriculture and Landscape Architecture,
HC & RI, TNAU, Coimbatore (Tamil Nadu), India.
²Associate Professor and Head, Grapes Research Station, Anaimalayanpatty, Theni, India.
³Professor (Horticulture), Department of Floriculture and Landscape Architecture,
HC & RI, TNAU, Coimbatore (Tamil Nadu) India.
⁴Professor (SS & AC), Department of Soil Science & Agricultural Chemistry,
TNAU, Coimbatore (Tamil Nadu) India.
⁵Assistant Professor (Crop Physiology), Department of Fruit Science,
HC & RI, TNAU, Coimbatore (Tamil Nadu) India.

(Corresponding author: A. Subbiah*) (Received: 13 June 2023; Revised: 25 June 2023; Accepted: 28 July 2023; Published: 15 August 2023) (Published by Research Trend)

ABSTRACT: African marigold (*Tagetes erecta* L.) is an important traditional flower crop under cultivation throughout India. It belongs to the family Asteraceae/Compositae. It has a great economic potential as loose flower which finds industrial application in preparation of natural dyes, essential oils and also used as nematode and mosquito repellents. The current investigation on "Impact of water soluble fertilizers and silica on the growth of African marigold (*Tagetes erecta* L.)" for the hybrid Tennis ball orange was carried out at Grapes Research Station, Theni. The experiment was laid out in Randomized Block Design (RBD) with fifteen treatments replicated thrice. The results revealed that application of the treatment T₂ [Mono Potassium Phosphate (0:52:34) @ 0.5% + Potassium Nitrate (13:0:45) - @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of Silica @ 0.1% + Norwegian sea weed extract (*Ascophyllum nodosum*) @ 0.1%] resulted in the maximum growth. The highest values for vegetative parameters *viz.*, plant height, internodal length, leaf length, leaf width, number of branches and stem diameter were recorded in the treatment T₂ followed by T₇ [Mono Potassium Phosphate (0:52:34) @ 0.5% + Potassium Nitrate (13:0:45) - @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of Silica @ 0.1%] when compared to the control (T₁₅).

Keywords: Tagetes erecta L., Tennis ball orange, water soluble fertilizers, ortho silicic acid and Ascophyllum nodosum.

INTRODUCTION

The African marigold (*Tagetes erecta* L.), which belongs to the Asteraceae/Compositae family, is a popular loose flower among flower cultivation. It is cultivated commercially throughout the India, including Tamil Nadu and it has gained increasing popularity among flower gardeners because of its ease in cultivation and adaptabilities (Patokar *et al.*, 2017). Marigold farming covers 84.09 thousand hectares in India with a production of 916.24 thousand tonnes of loose flowers during 2021-22. Whereas in Tamil Nadu, it occupies 3.39 000 hectares of area with the loose flower production of 96.31 thousand MT during 2021-22 (NHB Data Base, 2022).

Foliar fertilization is an effective technique for proper absorption that may be adapted to meet the plants with specific needs for one or more macro nutrients like Nitrogen (N), Phosphorous (P) and Potassium (K) and micro nutrients, particularly trace minerals and can be

used to correct deficiencies, strengthen, weak or damaged crops, quicken growth and make plants grow better (Said-Al Ahl and Mahmoud 2010). Meanwhile, other than macro and micro nutrients application of silica and seaweed extracts showed the greater effects in the growth and development of African marigold. Therefore, the present investigation has been formulated to standardize the vegetative growth of African marigold.

MATERIAL AND METHODS

The experiment was conducted in a Randomized Block Design (RBD) with 15 treatments combinations and replicated thrice at Grapes Research Station, Theni during the year 2023. Healthy and uniform seedlings of twenty one days old marigold hybrid - Tennis Ball Orange were collected from nursery and transplanted in ridges and furrows by adopting a spacing of 60 cm \times 45 cm in the open field condition.

The treatments include T₁ - Water soluble NPK mixture **@** 0.5% + Mono ammonium phosphate(12:61:0) @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of silica @ 0.1%, T₂ -Water soluble NPK mixture - 19:19:19 @ 0.5% + Mono ammonium phosphate (12:61:0) @ 0.5% + Chelated micronutrient @ 0.2% + Ortho silicic acid form of silica @ 0.2%, T₃ - Mono ammonium phosphate (12:61:0) @ 0.5% + Mono potassium phosphate (0:52:34) @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of silica @ 0.1%, T₄ -Mono ammonium phosphate (12:61:0) @ 0.5% + Mono potassium phosphate (0:52:34) @ 0.5% + Chelated micronutrient @ 0.2% + Ortho silicic acid form of silica @ 0.2%, T₅ - Mono Ammonium Phosphate (12:61:0) @ 0.5% + Mono potassium phosphate (0:52:34) @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of silica @ 0.1% + Norwegian sea weed extract (Ascophyllum nodosum) @ 0.1%, T₆ -Mono ammonium phosphate (12:61:0) @ 0.5% + Mono potassium phosphate (0:52:34) @ 0.5% + Chelated micronutrient @ 0.2% + Ortho silicic acid form of silica @ 0.2% + Norwegian sea weed extract (Ascophyllum nodosum) @ 0.2%, T₇ - Mono potassium phosphate (0:52:34) @ 0.5% + Potassium Nitrate (13:0:45) - @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of silica @ 0.1%, T₈ - Mono potassium phosphate (0:52:34) @ 0.5% + Potassium Nitrate (13:0:45) - @ 0.5% + Chelated micronutrient @ 0.2% + Ortho silicic acid form of silica @ 0.2%, T₉ -Mono potassium phosphate (0:52:34) @ 0.5% + Potassium Nitrate (13:0:45) - @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of silica @ 0.1% + Norwegian sea weed extract (Ascophyllum nodosum) @ 0.1%, T_{10} - Mono potassium phosphate (0:52:34) @ 0.5% + Potassium Nitrate (13:0:45) - @ 0.5% + Chelated micronutrient @ 0.2% + Ortho silicic acid form of silica @ 0.2% + Norwegian sea weed extract (Ascophyllum nodosum) @ 0.2%, T₁₁ - Mono potassium phosphate (0:52:34) @ 0.5% + Potassium Nitrate (13:0:45) - @ 0.5% + Potassium schoenite @ 0.2% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of silica @ 0.1%, T₁₂ - Mono potassium phosphate (0:52:34) @ 0.5% + Potassium Nitrate (13:0:45) - @ 0.5% + Potassium schoenite @ 0.2% + Chelated micronutrient @ 0.2% + Ortho silicic acid form of silica @ 0.2%, T₁₃ - Mono potassium phosphate (0:52:34) @ 0.3% + Potassium Nitrate (13:0:45) - @ 0.3% + Potassium schoenite @ 0.2% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of silica @ 0.1% + Norwegian sea weed extract (Ascophyllum nodosum) @ 0.1%, T₁₄ - Mono potassium phosphate (0:52:34) @ 0.3% + Potassium Nitrate (13:0:45) - @ 0.3% + Potassium schoenite @ 0.2% + Chelated micronutrient @ 0.2% + Ortho silicic acid form of silica @ 0.2% + Norwegian sea weed extract (Ascophyllum nodosum) @ 0.2%, T₁₅ - Control (No spray) in 14 different combinations were sprayed as foliar spray on 20 to 25 DAT, 40 to 45 DAT, 60 to 65 DAT and 80 to 85 DAT along with water spray as control. The vegetative parameters like plant height,

internodal length, leaf length, leaf width, number of branches and stem diameter were recorded at 15, 30, 45 and 60 days after planting (DAP). The data recorded were subjected to statistical analysis using AGRES software and MS Excel® spread sheet.

RESULTS AND DISCUSSION

The data of this investigation showed that growth parameters were positively influenced by foliar application. The plant height, the number of branches, leaf length, leaf width, number of leaves, stem diameter and internodal length, the fresh and dry weight of shoots and leaf area increased significantly by foliar application of water soluble fertilizers, chelated micronutrients, silica and sea weed extract.

A. Plant height

The treatment T₉ [Mono Potassium Phosphate (0:52:34) @ 0.5% + Potassium Nitrate (13:0:45) - @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of Silica @ 0.1% + Norwegian sea weed extract (Ascophyllum nodosum] @ 0.1%) envisaged the highest plant height (41 cm at 30 DAT (Fig. 1) and 56 cm at 45 DAT) than all other treatments followed by T₇ [Mono Potassium Phosphate(0:52:34) @ 0.5%+ Potassium Nitrate (13:0:45) - @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of Silica @ 0.1%] with 38 cm at 30 DAT and 52 cm at 45 DAT and T₅ [Mono Ammonium Phosphate (12:61:0) @ 0.5% + Mono Potassium Phosphate (0:52:34) @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of Silica @ 0.1% + Norwegian sea weed extract (Ascophyllum nodosum) @ 0.1%] with 36 cm at DAT and 51.67 cm at 45 DAT both are statistically on par with each other. The least plant height was recorded in the T₁₅ (control) at 30 DAT (Fig. 2) and T₃ [Mono Ammonium Phosphate (12:61:0) @ 0.5% + Mono Potassium Phosphate (0:52:34) @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of Silica @ 0.1%] at 45 DAT. Similar results were reported by Shyala et al., (2019).

B. Number of branches

Number of branches shows the significance difference between all the treatments and the highest number of branches was recorded in the treatment T9[Mono Potassium Phosphate (0:52:34) @ 0.5% + Potassium Nitrate (13:0:45) - @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of Silica @ 0.1% + Norwegian sea weed extract (Ascophyllum nodosum) @ 0.1%] with 32.39 Nos at 45 DAT and least number of branches was recorded in treatment T2 [Water soluble NPK mixture - 19:19:19 @ 0.5% + Mono Ammonium Phosphate (12:61:0) @ 0.5% + Chelated micronutrient @ 0.2% + Ortho silicic acid form of Silica @ 0.2%] with 24.76 Nos. Increased number of branches could be due to micronutrients like ZnSO₄ and FeSO₄, which are essential components of several dehydrogenase, proteinase, peptidase and promote hormone growth and are closely associated with growth. All of these factors contributed to cell multiplication, cell division and cell differentiation, resulting in increased photosynthesis and translocation of food material, which increased the number of branches, above result was confirmed by Pal *et al.* (2016). Similar results were recorded by Kumar and Moon (2014); Polara *et al.* (2015).

C. Leaf length and width

Leaf length shows significantly variation among the treatments and leaf width shows non-significant at 30 DAT and shows significant at 45 DAT. The treatment T₉ [Mono Potassium Phosphate (0:52:34) @ 0.5% + Potassium Nitrate (13:0:45) - @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of Silica @ 0.1% + Norwegian sea weed extract (Ascophyllum nodosum) @ 0.1%] recorded the highest leaf length (3.55 cm at 30 DAT and 4.55 cm at 45 DAT) than all other treatments followed by T7 [Mono Potassium Phosphate(0:52:34) @ 0.5%+ Potassium Nitrate (13:0:45) - @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of Silica @ 0.1%] with 3.43 cm at 30 DAT and 4.35 cm at 45 DAT and the least leaf length was recorded in the treatment T₆ [Mono Ammonium Phosphate (12:61:0) @ 0.5% + Mono Potassium Phosphate (0:52:34) @ 0.5%+ Chelated micronutrient @ 0.2% + Ortho silicic acid form of Silica @ 0.2% + Norwegian sea weed extract (Ascophyllum nodosum) @ 0.2%] at 45 DAT and the least leaf width was recorded in the treatment T₁₀ Phosphate(0:52:34) Mono Potassium 0.5%+Potassium Nitrate (13:0:45) - @ 0.5% + Chelated micronutrient @ 0.2% + Ortho silicic acid form of Silica @ 0.2% + Norwegian sea weed extract (Ascophyllum nodosum) @ 0.2%] at 45 DAT.

DISCUSSION

Fertilizing a plant through foliar spray can make plant to quickly absorb the nutrients and reduce the usage of bulk fertilizers. Water soluble fertilizers are easily adopted to this methodology. Potassium plays an important role in protein and starch synthesis which is responsible for vegetative growth and nitrogen increases the meristematic activity and amino acid formation has significant effect on plant growth (Dali et al., 2019). Phosphorus treatment influences the plant spread and total leaf chlorophyll content, but had little effect on other plant growth indices by Polara et al., (2015) and Shah et al., (2018). Micronutrients stimulate many enzymes (catalase, carbonic dehydrogenase and tryptophan synthases etc.) and plays a vital role in a various physiological functions. Zn enhances cell multiplication, division, and differentiation, which results in higher photosynthesis and food material translocation, which increases plant (Thirumalmurugan et al., 2021). By the application of micronutrients, Kakade et al. (2009) reported that spraying of ZnSO₄ produces maximum vegetative growth in China aster and similarly, Balakrishnan (2005) reported in marigold, Pradhan & Mitra (2020) in marigold. Many of the studies emphasised the importance of silicon in plant resilience to both biotic and abiotic stresses including salinity (Crusciol et al., 2009). Many researchers also confirms that, by silicon foliar spray, plants can withstand long periods of drought conditions and also increases flower shelf life.

Table 1: Impact of water soluble fertilisers and silica on the growth of African marigold after 15 DAT.

	15 Days After Transplanting						
Treatments	Plant height (cm)	Internodal length (cm)	Leaf length (cm)	Leaf width (cm)	No. of branches per plant	Stem diameter (cm)	
T_1	20.00	2.53	2.60	0.45	13.85	1.23	
T ₂	17.46	2.40	2.58	0.40	13.05	1.10	
T ₃	19.40	2.43	2.70	0.40	12.05	1.13	
T ₄	18.18	2.37	2.65	0.37	13.25	1.07	
T ₅	18.05	2.47	2.68	0.40	13.95	1.10	
T ₆	18.43	2.33	2.57	0.35	12.45	1.20	
T ₇	21.63	2.60	2.82	0.47	15.34	1.33	
T ₈	20.00	2.37	2.65	0.35	12.21	1.20	
T9	22.00	2.83	2.92	0.52	18.75	1.43	
T ₁₀	18.43	2.37	2.65	0.35	10.27	1.10	
T ₁₁	17.46	2.37	2.72	0.37	12.37	1.20	
T ₁₂	19.00	2.40	2.65	0.38	11.58	1.13	
T ₁₃	17.46	2.37	2.70	0.37	13.24	1.17	
T ₁₄	19.40	2.37	2.62	0.40	12.62	1.20	
T ₁₅	19.19	2.37	2.65	0.35	13.71	1.20	
Mean	19.33	2.44	2.7	0.49	13.25	1.19	
SE (d)	0.43	0.07	0.06	0.53	0.30	0.05	
CD (0.05)	0.87	0.15	0.12	1.08	0.61	0.10	
	**	**	**	NS	**	**	

^{(** -} Significance, NS – Non-significance)

Table 2: Impact of water soluble fertilisers and silica on the growth of African marigold after 30 DAT.

		30 Day	s After Transplanting				
Treatments	Plant height (cm)	Internodal length (cm)	Leaf length (cm)	Leaf width (cm)	No. of branches per plant	Stem diameter (cm)	
T_1	37.33	3.30	3.23	0.65	22.67	2.73	
T_2	36.00	3.20	3.23	0.68	21.43	2.60	
T_3	36.00	3.23	3.37	0.72	21.78	2.60	
T_4	36.67	3.27	3.35	0.67	22.76	2.53	
T_5	37.33	3.30	3.34	0.70	22.83	2.57	
T_6	36.00	3.23	3.27	0.63	22.91	2.70	
T_7	38.00	3.40	3.43	0.82	24.62	2.83	
T_8	36.67	3.30	3.27	0.70	23.27	2.70	
T ₉	41.00	3.48	3.55	1.07	26.87	2.93	
T_{10}	36.00	3.20	3.23	0.70	22.08	2.60	
T_{11}	36.00	3.30	3.27	0.70	22.61	2.70	
T_{12}	37.33	3.30	3.40	0.73	23.08	2.63	
T_{13}	36.00	3.28	3.25	0.72	22.68	2.67	
T_{14}	36.67	3.28	3.28	0.69	22.91	2.70	
T_{15}	36.00	3.28	3.27	0.69	22.63	2.70	
Mean	36.8660	3.2896	3.3167	0.8091	23.0087	2.6800	
SE (d)	0.8054	0.8054	0.0723	0.4594	0.5058	0.0583	
CD (0.05)	1.6498	1.6498	0.1481	0.9410	1.0362	0.1194	
	**	*	**	NS	**	**	

^{(** -} Significance, NS – Non-significance)

Table 3: Impact of water soluble fertilisers and silica on the growth of African marigold after 45 DAT.

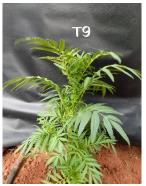
45 Days After Transplanting						
Treatments	Plant height (cm)	Internodal length (cm)	Leaf length (cm)	Leaf width (cm)	No. of branches	Stem diameter (cm)
T_1	49.67	3.83	4.17	1.13	27.42	3.10
T_2	49.67	3.77	4.10	1.06	24.76	3.23
T_3	44.33	3.70	4.07	1.05	28.08	3.40
T_4	48.00	3.77	4.00	1.05	25.71	3.32
T ₅	51.67	3.83	4.13	1.00	28.04	3.45
T ₆	49.67	3.63	3.97	1.07	26.75	3.35
T ₇	52.00	3.87	4.35	1.14	29.89	3.72
T_8	49.67	3.70	4.07	1.03	25.68	3.54
T ₉	56.00	3.93	4.55	1.25	32.31	3.98
T_{10}	48.00	3.70	4.00	0.98	26.81	3.43
T ₁₁	50.00	3.70	4.15	1.00	27.03	3.54
T ₁₂	49.67	3.68	4.13	1.02	28.01	3.32
T ₁₃	49.00	3.70	4.22	1.07	27.98	3.45
T ₁₄	49.00	3.73	4.12	1.01	26.88	3.48
T ₁₅	48.00	3.73	4.03	1.02	26.64	3.44
Mean	49.6227	3.7518	4.1371	1.0642	27.4660	3.4500
SE (d)	1.0929	0.0826	0.0826	0.0689	0.6075	0.0759
CD (0.05)	2.2387	0.1691	0.1859	0.1411	1.2445	0.1555
	**	NS	**	**	**	**

^{(** -} Significance, NS – Non-significance)

Table 4: Impact of water soluble fertilisers and silica on the dry matter production and Leaf area of African marigold.

Treatments	Fresh weight of leaf (g)	Dry weight of leaf (g)	Leaf area (cm)
T_1	1.50	1.09	27.61
T_2	1.57	1.11	28.43
T_3	1.55	1.07	27.18
T_4	1.56	1.09	29.31
T_5	1.43	1.06	26.43
T_6	1.30	1.06	27.68
T_7	1.67	1.12	31.90
T_8	1.40	1.06	28.67
T ₉	1.75	1.15	34.15
T_{10}	1.50	1.07	26.45
T_{11}	1.45	1.08	29.38
T ₁₂	1.59	1.08	29.58
T ₁₃	1.60	1.07	30.20
T_{14}	1.58	1.06	28.38
T ₁₅	1.56	1.05	25.65
Mean	1.53	1.08	28.7336
SE (d)	0.0341	0.0221	0.6274
CD (0.05)	0.0698	0.0452	1.2853
	**	**	**

 $^{(** \}hbox{ - Significance}, NS-Non\hbox{--significance})$



Treatment 9 - 30 DAT **Fig. 1.**



Treatment 15 - 30 DAT **Fig. 2.**

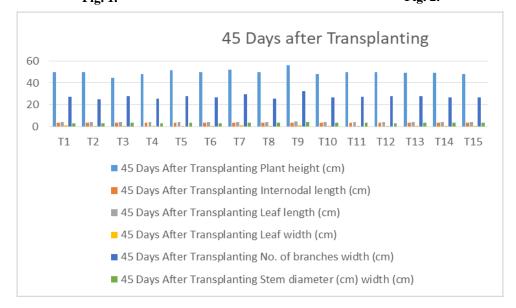


Fig. 3. Impact of water soluble fertilizers and silica on the growth of African marigold after 45 DAT.

CONCLUSIONS

The study concluded that application of the treatment T_9 (Mono Potassium Phosphate (0:52:34) @ 0.5% + Potassium Nitrate (13:0:45) - @ 0.5% + Chelated micronutrient @ 0.1% + Ortho silicic acid form of Silica @ 0.1% + Norwegian sea weed extract (*Ascophyllum nodosum*) @ 0.1%) recorded maximum growth with the highest values for vegetative parameters viz., plant height, internodal length, leaf length, leaf width, number of branches and stem diameter when compared to the control (T_{15}).

FUTURE SCOPE

The main aim of this study is to access the efficiency of fertilizers through foliar application and thereby reducing the usage of bulk quantities of fertilizers which also increases benefit cost ratio to farmers and have practical benefit.

Acknowledgement. I would proudly take this opportunity to express my deep sense of thanks and gratitude to my chairperson Dr. A. Subbiah for his valuable guidance and support. Also I owe to the company M/s. Agri. Search India Ltd., Nashik, Maharashtra, which helped me by providing the funds and materials throughout my research work. I extremely grateful to members of my advisory committee for their

advice throughout my studies. Finally I feel blessed to dedicate this to my loveable Grandparents.

REFERENCES

Balakrishnan, V. (2005). Effect of micronutrients on flower yield and xanthophylls content of African marigold (*Tagetes erecta* L.). M.Sc., (Hort.), Thesis submitted to Tamil Nadu Agricultural University, Coimbatore.

Crusciol, C. A., Pulz, A. L., Lemos, L. B., Soratto, R. P., & Lima, G. P. (2009). Effects of silicon and drought stress on tuber yield and leaf biochemical characteristics in potato. *Crop science*, 49(3), 949-954.

Dali, N. M., Khobragade, Y. R., Vasu, A. S., Gajbhiye, R. P., & Panchbhai, D. M. (2019). Assessment of nitrogen and potassium levels for growth, flowering and yield attributes in African marigold. *Journal of Pharmacognosy and Phytochemistry*, 8(5), 1296-1299.

Kakade, D. K., Rajput, S. G., & Joshi, K. L. (2009). Effect of foliar application of Fe and Zn on growth, flowering and yield of China aster (*Callistepus chinensis L. Nees*). *Asian Journal of Horticulture*, 4(1), 138-140.

Kumar, N. V., & Moon, S. S. (2014). Effect of phosphorus and potassium on growth and flowering of African marigold. *Journal of Soils and Crops*, 24(1), 169-173.

Pal, S., Barad, A. V., Singh, A. K., Khadda, B. S. & Kumar D. (2016). Effect of foliar application of Fe and Zn on growth, flowering and yield of gerbera (*Gerbera jamesonii*) under protected condition. *Indian J Agri. Sci.*, 86(3), 394-398.

Patokar, M. J., Chopde, N., & Kuchanwar, O. (2017). Effect of micronutrients (Zn and Fe) as a foliar spray on growth

- and flower production of marigold. *Plant Archives*, 17(1), 312-314
- Polara, N. D., Gajipara, N. N., & Barad, A. V. (2015). Effect of nitrogen and phosphorus nutrition on growth, flowering, flower yield and chlorophyll content of different varieties of African marigold (*Tagetes erecta L.*). Journal of Applied Horticulture, 17(1), 44-47.
- Pradhan, S., & Mitra, M. (2020). Effect of Micronutrients on Growth and Flowering of Marigold cv. Siracole. Agricultural Science Digest-A Research Journal, 40(1), 53-56.
- Said-Al Ahl, H. A. H., & Mahmoud, A. A. (2010). Effect of zinc and/or iron foliar application on growth and essential oil of sweet basil (*Ocimum basilicum* L.) under salt stress. *Ozean Journal of Applied Sciences*, 3(1), 97-111.
- Shah, F. A., Khan, T., Ahmad, I., Shahid, M. A., Khan, S., & Ibrahim, M. (2018). Response of marigold (*Tagetes erecta* L.) to different levels of nitrogen at Bagh E Naran Park Peshawar. *International Journal of Environmental Sciences & Natural Resources*, 14(1), 01-03.
- Shyala, M. R., Dhanasekaran, D., & Rameshkumar, S. (2019). Effect of foliar application of micronutrients and potassium humate on growth and flower yield of African marigold (*Tagetes erecta L.*). Annals of Plant and Soil Research, 21(2), 101-107.
- Thirumalmurugan, V., Manivannan, K., & Nanthakumar, S. (2021). Influence of micronutrients on growth, flowering and yield of African marigold (*Tagetes erecta* L.). *Journal of Pharmacognosy and Phytochemistry*, 10(3), 461-463.

How to cite this article: Bharathkumar C., A. Subbiah, D. Keisar Lourdusamy, K. Sathiya Bama and K. Vanitha (2023). Impact of Water Soluble Fertilizers and Silica on the Growth of African Marigold (*Tagetes erecta* L.). *Biological Forum – An International Journal*, 15(8): 561-566.