

Scientific Investigation of *Pavonia zeylanica* (L.) Cav. A Potent Medicinal Plant

S.M. Dhivya^{1*}, P. Vijayashalini¹, P. Abirami² and S. Sharmila¹

¹Assistant Professor, PG and Research Department of Botany,
Vellalar College for Women (Autonomous), Thindal, Erode (Tamil Nadu), India.

²Associate Professor & Head, PG and Research Department of Botany,
Vellalar College for Women (Autonomous), Thindal, Erode (Tamil Nadu), India.

(Corresponding author: S.M. Dhivya*)

(Received: 15 August 2024; Revised: 14 September 2024; Accepted: 11 October 2024; Published: 14 November 2024)

(Published by Research Trend)

ABSTRACT: The examination was carried out to establish the possible bioactive components of entire plant of *Pavonia zeylanica* by Qualitative investigation, UV –VIS, FT-IR and GC-MS analysis. FT-IR spectroscopy is a well-established and efficient method for characterizing and identifying functional groups in the ethanol extract of the entire *Pavonia zeylanica* plant. At Vellore Institute of Technology (VIT), GC-MS analysis was conducted to determine the chemical composition of some of the powerful volatile constituents present in the extract. The ethanol extract showed the positive result to alkaloids, phenols, flavonoids, quinone, coumarin, gum and mucilage, fixed oil & fat. The UV-VIS Spectrum profile showed the peaks at 245, 743 and 665 nm with the absorption of 0.106, 0.006, and 0.006 correspondingly. The FT-IR showed a broad peak at 3.398.57 cm⁻¹ which indicated the presence of N-H stretch and may be attributed to amine and the peak around 1427.32 cm⁻¹ are due to carbonate group, peak at 1265.3 cm⁻¹ represents hydroxyl O-H bending. The FT-IR spectrum confirmed the occurrence of alkane, haloalkane, hydroxyl, nitrile, and amine in powder pellet. Fifteen chemical substances belonging to diverse categories were identified. The peak area percentage of N-Hexadecanoic acid (23.280%) was found to be maximum when compared to other compounds. The study comes to the conclusion that the species *Pavonia zeylanica* may provide bioactive substances such as amines, ketone, alcohols, esters, and alkenes, among others. The traditional use of this species is justified by this study.

Keywords: GC-MS, *Pavonia zeylanica*, phytol, ketone and esters.

INTRODUCTION

Both conventional and modern medical procedures constantly emphasise the importance of plants. Plants are primarily useful as medicines because they contain a variety of phytoconstituents, including tannins, flavonoids, phenolic chemicals, and alkaloids. Phytochemicals are non-nutritive, bioactive compounds obtained from plants that are good for human health and disease prevention. These compounds give plants their distinctive scent and colour and are crucial to their defence mechanisms against illness (Bano and Deora 2019).

There is much more to medicinal plant research than just finding new medications. Natural products, moreover as unadulterated compounds or as standardized plant extracts, offer limitless opportunities for new drug. This field has been expanding and includes various subjects as negotiation of value based on medicinal plant knowledge. The diverse phytoconstituents present in plants contain arthra glycosides, bitter drugs, flavonoids, alkaloids, saponins, coumarins, phenol carboxylic acids and terpenes. Spectroscopic (UV-Vis, FT-IR) methods jointly or separate can be used in this sense as well as conventional methods (Dhivya and Kalaichelvi 2017).

In this study the medicinal plant *Pavonia zeylanica* belonging to the family Malvaceae was taken for scientific evaluation. The genus *Pavonia* includes around 30 species. The plant is origin of Eurasia and North Africa, but later migrated to all continents except the two poles. Numerous review articles have been published about the conventional uses of *Pavonia* species, but to the best of our knowledge, there are no published review articles that summarize the scientific literature regarding all *Pavonia* species surveyed (Abdullatif Azab, 2017).

Phytochemicals show tremendous potential in the prevention and treatment of wounds and microbial infections (Ayaz *et al.*, 2015; 2017). Antimicrobial, antioxidant, and wound-healing phytochemicals promote blood coagulation, combat infection, and hasten wound healing. Medicinal plants wealthy in polyphenols are reported to acquire notable anti-oxidant activity (Ovais *et al.*, 2018; Mehta *et al.*, 2015; Zohra *et al.*, 2019). Phenolics encourage wound healing chiefly due to their astringent, antimicrobial and free radical scavenging properties (Deshmukh *et al.*, 2009; Lopes *et al.*, 2005). Finally, polyphenolic components like flavonoids can encourage tremendous curative of wounds probably by means of antimicrobial and anti-

oxidative property, in that way inhibiting the lipid peroxidation, which led to the avoidance of cell damage and increase in the viability of collagen fibrils (Getie *et al.*, 2002; Shetty *et al.*, 2008).

Phytochemical screening is vital for validating the conventional use of therapeutic plants. In addition GC-MS analysis method can be used to study traditional remedy and to exemplify the compounds of interest. Ultraviolet-visible spectrophotometry (UV-Vis) related to the spectroscopy of photons in the UV-visible region. UV-visible spectroscopy uses light in the visible ranges or its nearby ranges. The colour of the chemicals concerned directly affects the absorption in the visible ranges. Molecules undergo electronic transitions in these ranges of the electromagnetic spectrum (Dhivya and Kalaichelvi 2017). The Fourier Transform Infrared spectroscopy (FT-IR) allows the analysis of a applicable quantity of compositional and structural information in plants. Furthermore, FT-IR spectroscopy is an recognized time-saving process to characterize and identified functional groups.

Analytical method that gives an impression of a tissue's metabolic makeup at a specific moment but does not resolve the concentrations of particular metabolites (Griffiths and De Haseth 1986). The structure of an unknown composition and the strength of the absorption spectra linked to the molecular composition or concentration of the chemical group can be determined using FT-IR. The vibrations of bonds within chemical functional groups are measured by the FT-IR method, which produces a spectrum that can be thought of as a biological sample. The concrete structure of some plant secondary metabolites has currently been identified using FT-IR, especially in phytochemistry (Yang and Yen 2002; Ivanova and Singh 2003). But, on pharmacognosy FT-IR is still a novel tool to characterize and recognize the trade components from the adulterant. FT-IR method has been effectively utilized in the characterization of bacterial, fungal and plant species. FT-IR is one of the most extensively used methods to identify the chemical constituents and elucidate the compounds structures, and has been used as a necessary technique to identify medicines in Pharmacopoeia of several countries. During last few decades UV-VIS, FT-IR and GC-MS were acted as influential techniques for the recognition, separation and structural determination of phytochemicals. Gas chromatography- mass spectroscopy (GC-MS) helped in identification of compounds at less than 1 mg. usually GC-MS applied for drug finding, environmental research and discovery of unknown samples. GC-MS method has been founds very effective for the parting and detection of composite mixtures of phytochemicals. UV-Visible and FT-IR can be used collectively or separately as conventional methods to identify phytoconstituents (Prabhu and Ramar 2018).

MATERIALS AND METHODS

Pavonia zeylanica (L.) Cav.

Collection of Specimen. The leaves of *Pavonia zeylanica* was gathered from Sulakarai, Krishnagiri Dhivya *et al.*,

district, Tamil Nadu, India. The plant material was dried separately under shade and pulverized in a motorized chopper and stored in a closed container for future use.

Plant Description. Its large, branching herb can grow to a height of 1-1.5 m. On flower stalks and stem leaves, hairs have been found. Leaves are 1.5-3cm long, 1-2.5 cm broad and lance-shaped to ovate. However, subordinate leaves are 3-lobed and lobes oblong or obovate. Leaf stalks are of 1-4.5 cm long. Flowers are originate singly in leaf axils and are about 1.5 cm long and pink in colour. The length of flower stalk is about 2-4 cm long. Sepals are lance-shaped. Fruit is velvety, spherical and about 5 mm. This shrub is mainly found in the countries like Srilanka, India, Pakistan, Arabia and Tropical Africa (Joga Rao *et al.*, 2020) Plate 1.

Traditional Medicinal uses. The plant leaves are macerated and paste, the paste is applied over the broken limbs for 1-2 weeks to cure bone fracture. The leaves are used for the treatment of Ezema (Perumal Samy and Ignacimuthu 2000). The stem and leaves of *Pavonia zeylanica* has an analgesic and anti-inflammatory property (Hepey Kalarani *et al.*, 2012). The root portion is thoroughly washed and boiled to prepare a decoction. This preparation is thoroughly sieved and used to control dysentery and abdominal pain. Another remedy for reducing itching is the root decoction combined with turmeric. Fresh leaf paste is used as a treatment on wounds to reduce inflammation and other skin problems. (Joga Rao *et al.*, 2020).

Shade drying and powdering of the collected Specimens. Fresh leaves of experimental plant were cleaned to eliminate adhering dirt and then shade dried. For phytochemical analysis, the shade-dried plant materials were mechanically ground into a coarse powder and put through a Willy Mill to achieve a 60-Mesh size. Samples were kept in high-quality plastic containers and kept at room temperature until analysis (Harborne, 1973).

Extraction procedure: Using a hot percolation method, benzene, ethanol, and water were used to extract coarsely powdered plant material using a Soxhlet apparatus. After that, the extracts were used to test the preliminary phytochemicals.

Phytochemical analysis

Qualitative phytochemical analysis. Phytochemical analyses are done to provide common plan about the nature of constituents present in the crude extract. The phytochemical screening of constituents *viz.*, carbohydrates, proteins, alkaloids, aminoacids, flavonoids, tannins, phenols, terpenoids, steroids, saponins, coumarin, quinine, anthraquinone, glycosides, gum and fixed oil present in the leaves of *Pavonia zeylanica* in various extracts (Benzene, Ethanol and aqueous) were carried out by the following standard procedures of Harborne (1998).

Plant extracts preparation for FT-IR and UV-VIS studies. The shade dried leaves of *Pavonia zeylanica* (at 25°C) was powdered in a mechanical grinder. 20 gms of leaves powder was weighed; 150 ml of Ethanol was added and retained for 3 days. Whatman No. 1

filter paper was used to filter the extract, and the supernatant was then gathered. Two further extractions of the residue were performed (with 3 days of the interval for each extraction) and the supernatants were collected. Supernatants were pooled and evaporated (at room temperature, 28 ± 1 °C) until the quantity was condensed to 150 ml. Extract of the entire plant powder with ethanol was prepared and kept in airtight bottles for after examination.

UV-VIS spectrum examination. The extract was centrifuged at 3000 rpm for 10 min and filtered through Whatman No.1 filter paper. The sample was diluted to 1:10 with the similar solvent. The extract was scanned at wavelength range from 200 to 1100 nm by Perkin Elmer Spectrophotometer and the characteristic peaks were detected. The peak values of the UV-VIS were recorded.

FT-IR analysis. Dried powder (ethanolic extract) of test plant was used for FT-IR analysis. 1 mg of the dried powder was encapsulated in 10 mg of KBr pellet, in order to prepare translucent sample discs. The powdered sample of the pellet was loaded in FT-IR spectroscopy (Shimadzu, Japan), with a Scan range from 400 to 4000cm^{-1} with a resolution of 4cm^{-1} .

Gas chromatography-Mass spectrum (GC-MS). Ethanolic extract of plant sample was analyzed for the occurrence of diverse volatile compounds by Gas Chromatography-Mass Spectroscopy (GC-MS) method. GC-MS analysis of a little of the potent volatile constituents present in the extract was performed at Vellore Institute of Technology (VIT), Vellore, Tamil Nadu, India. GC analysis of the extracts was performed using a GC-MS (Model; Thermo Trace GC Ultra Ver.5.0, Thermo MS DSQ II) equipped with a DB-35MS capillary standard Non – polar column (30m length X outside diameter 0.25 mm X internal diameter 0.25 μm) and gas chromatograph interfaced to a Mass Selective Detector (MS-DSQ-II) with XCALIBUR software. GC analysis was carried out at a Clarus 500, Perkin Elmer and Computer MS Programme Library used NIST Version -2005. The capillary column was Elite-5MS (5% Diphenyl/95% Dimethyl poly siloxane), $30 \times 0.25\text{mm} \times 0.25\mu\text{m}$ df. The mass detector was Turbo mass gold-Perkin Elmer. The extract was injected in the split mode with 10:1 ratio, 1ml per minute.

For GC-MS detection, an electron ionization system with ionization energy of -70eV was used. Helium gas was used as a carrier gas at a constant flow rate of 1ml/min and the sample injected was 1 μl ; Injector temperature 260°C; Ion source temperature 200°C. The oven temperature was programmed from 75°C to 260°C at the rate of 10°C/min, held isothermal for 1 minute and finally raised to 260°C at 6°C/min. Interface temperature was kept at 260°C. Total GC run time was 37.53 min. The relative percentage of each extract constituent was expressed as percentage with peak area normalization.

The components were identified after comparison with those available in the computer library attached to the

GS-MS instrument and reported in literature (Jennings and Shibamoto 1980; Anonymous, 1962).

RESULTS AND DISCUSSION

Qualitative phytochemical screening. Successful evaluation of phytochemicals from plant material was mainly dependent on the kind of solvent used in the extraction method. The current work on the various solvent leaf extracts of *Pavonia zeylanica* shown the presence of alkaloids, tannin, phenols, flavonoids, terpenoid, saponin, quinone, coumarin, gum & mucilage, fixed oil & fat (Table 1). The Benzene leaf extract shows the presence of alkaloids, phenols, flavonoids, coumarin, fixed oil & fat. The same extract showed the negative result to carbohydrate, protein, aminoacids, tannin, terpenoid, saponin, glycoside, anthraquinone, quinone, gum & mucilage. The ethanol extract showed the positive result to alkaloids, flavonoids, coumarin, phenols, quinone, gum and mucilage, fixed oil & fat. The aqueous extracts revealed the occurrence of half of the compounds except carbohydrate, protein, aminoacid, glycoside, anthraquinone, fixed oil & fat. Aqueous extracts comprised greater secondary metabolites with a high degree of precipitation (+++) than other solvent extracts. The (+) or (-) symbol indicates whether an ingredient is present or absent.

The occurrence of phytochemical compounds like alkaloids, tannins, flavonoids and phenolic compounds in There are many uses for medicinal plants, and they might be applied in many different utilization (Lena, 2010). Our findings concur with the research of Edeoga *et al.* (2005). All of the test plants extract contained flavonoids, tannin, phenols, saponins, proteins, and amino acids. This is consistent with research of Leon Stephan Raj *et al.* (2015). The alkaloids present in plants were used in medicine as aesthetic agents (Leon Stephan Raj *et al.*, 2015). Presence of tannin in test plant could be useful in the treatment connected with heart, anti-inflammatory action, anticoagulant, diarrhoea and dysentery (Bokhad *et al.*, 2012). In the current study tannin were present in aqueous extract of the test plant.

Phenols were present in benzene, ethanol and aqueous extracts of all the test plant. Phenols and phenolic substances in the plants indicated that these plants may be used as an anti-microbial agent. This was in agreement with the result of Ofokansi *et al.* (2005). Saponins have the property of precipitating and coagulating red blood cells (Sodipo *et al.*, 2000; Okwu and Okwu 2004). Quinones were present in ethanol and aqueous extracts of the test plant. Quinones were used as adrenergic nervous system and hemostatic action (Ridhambaradevi *et al.*, 2018).

Benzene, ethanol, and aqueous extracts of the study plant contained coumarin. Due to their antibacterial properties, coumarins are also known to inhibit gram-positive bacteria and it is produced in carrots in response to fungal disease which could be attributed to its antimicrobial activity (Shihabudeen *et al.*, 2010).

When compared to synthetic compounds phytochemicals have low side effects and an excellent dose response. The studies conducted on leaves of *Pavonia zeylanica* showed the presences of phytochemicals. Their medicinal benefits are due to the existence of secondary metabolites called phytochemicals. It also expresses optimism for the creation of several more unique therapeutic agents or templates from these plants, which might one day be used to produce synthetically enhanced curative compounds.

UV-VIS spectrum and FT-IR study. Spectroscopic technique has developed into a potent and analytical tool for the qualitative and quantitative study of pharmaceutical and biological materials. Dried powder of *Pavonia zeylanica* was subjected to UV-VIS and FT-IR analysis. The qualitative UV-VIS spectrum report of ethanol extract of *Pavonia zeylanica* was selected at wavelength from 200 to 1100 nm due to sharpness of the peaks and proper baseline. The report showed the peaks at 245, 743 and 665 nm with the absorption of 0.106, 0.006, and 0.006 respectively (Table 2 & Fig. 1) and these bands represent the data for the occurrence of flavonoids.

The identification of secondary metabolite profiles by spectroscopy and chromatography yields important insights on the quantitative and qualitative makeup of plant species and how best to recognise them through chemometry. By differentiating between lipophilic and hydrophilic compounds based on polarity, UV-VIS spectroscopy provides an easy technique for identifying the primary phytochemicals. Spectroscopic (UV-VIS, FT-IR) methods jointly or separate can be used in this sense as well as in conventional methods.

The results of FT-IR peak values and functional groups are represented in Table 3 and the FT-IR spectrum profile is illustrated in Fig. 2. The FT-IR showed a broad peak at $3.398.57\text{ cm}^{-1}$ which indicated the existence of N-H stretching and may be attributed to amine. It displayed strong peaks at 2929.87 and 2362.8 cm^{-1} , which indicated the occurrence of O-H and $\text{C}\equiv\text{N}$ stretching vibrations corresponds the nitrile and alkane, peak at 1643.35 cm^{-1} and 1043.49 cm^{-1} attributed to C-F represents the haloalkane, the peak around 1427.32 cm^{-1} are due to carbonate group, peak at 1265.3 cm^{-1} represents hydroxyl O-H bending. There was no absorbance in between the region 2220 and 2260 cm^{-1} which indicated that there was no cyanide group in this extract. The present study revealed that *Pavonia zeylanica* does not have any lethal substances. The FT-

IR spectrum confirmed the existence of alkane, haloalkane, hydroxyl, nitrile, and amine in powder pellet.

Fourier Transform Infrared spectroscopy (FT-IR) is a high-resolution analytical method to identify the chemical constituents and clarify the structural compounds (Hashimoto and Kameoka, 2005; Hussain *et al.*, 2007), FT-IR offers a fast and non destructive study to fingerprint plant extracts or powders. In addition FT-IR spectroscopy is proved to be a consistent and sensitive technique for finding of biomolecular composition (Komalkumar and Devi Prasad 2011).

GC-MS analysis. The ethanol extract study plant was subjected to Gas Chromatogram and Mass Spectral (GC-MS) studies. Table 4 & Fig. 3 exhibited the presence of chemical constituents with their molecular formula, molecular weight, retention time, percentage of peak area, functional groups, biological activity, compound structure and hit spectrum. Fifteen chemical compounds belonging to diverse categories were recognized. Ethane,1,1-diethoxy- (7.376%), 4H-Pyran-4-one,2,3-dihydro-3,5-dihydroxy-6-methyl- (5.161%), 2-Methyl-6-methylene-octa-1,7-dien-3-ol (2.412%), 3-Tetradecen-5-yne,(E)- (3.330%), Trans-z.alpha.-bisabolene epoxide (1.797%), 3,7,11,15-Tetramethyl-2-hexadecen-1-ol (5.482%), 9-Methyl-z,z-10,12-hexadecadien-1-ol acetate (2.912%), N-Hexadecanoic acid (23.280%), Phytol (6.412%), 1-Tridecane (2.088%), 3-Tetradecyn-1-ol (11.767%), Pentadecanoic acid (5.291%), Oleic acid (2.984%), 6,11-Dimethyl-2,6,10-dodecatrien-1-ol (2.711%), Bisanallocholanic acid(2.613%) are the chief compounds present in the ethanol extract. The peak area percentage of N-Hexadecanoic acid (23.280%) was found to be maximum when compared to other compounds.

The chief components present in the flowers are 2,3-hexanediol (RT:17.59), n-Hexadecanoic acid (RT:26.02), 1,2- Benzenedicarboxylic acid (RT: 35.91) and squalene (RT: 41.55). GC-MS chromatogram of leaf methanolic extract of the plant *Abutilon pannosum* showed various peaks indicating presence of 50 phytochemical compounds. The principle components recognized with highest peak area was n-Hexadecanoic acid (13.38) at retention time 25.47 minutes and lowest peak area recorded was of 2-Butanone, 4-(2, 6, 6-trimethyl-1, 3cyclohexadien-1-yl)-(0.15) with retention time 16.97 minutes (Bano and Deora 2019).

Table 1: Preliminary phytochemical analysis of test plant.

Sr. No.	Metabolites	<i>Pavonia zeylanica</i>		
		Benzene	Ethanol	Aqueous
1.	Carbohydrate Test Barford's Test	-	-	-
2.	Protein Test Biuret Test	-	-	-
3.	Aminoacid Test Ninhydrin Test	-	-	-
4.	Alkaloids Wagner's Test	+	+	+

5.	Tannin Ferric Chloride Test	-	-	+
6.	Phenols Lead acetate Test	+	+	+
7.	Flavonoids Ammonium hydroxide Test	+	+	+
8.	Terpenoid Test	-	-	+
9.	Saponin Test Foam Test	-	-	+
10.	Glycoside Test Borntrager's Test	-	-	-
11.	Anthraquinone Test	-	-	-
12.	Quinone Test	-	+	+
13.	Coumarin Test	+	+	+
14.	Gum and Mucilage Test	-	+	+
15.	Fixed oil and fat Test	+	+	-

Table 2: UV-VIS Spectrum Peak values of ethanolic extracts of *Pavonia zeylanica*.

Sr. No.	Wavelength nm.	Abs.
1.	665.00	0.004
2.	466.00	-0.002
3.	337.00	0.041
4.	273.00	0.270
5.	743.00	-0.006
6.	583.00	-0.007
7.	457.00	-0.002
8.	305.00	0.029
9.	245.00	0.106

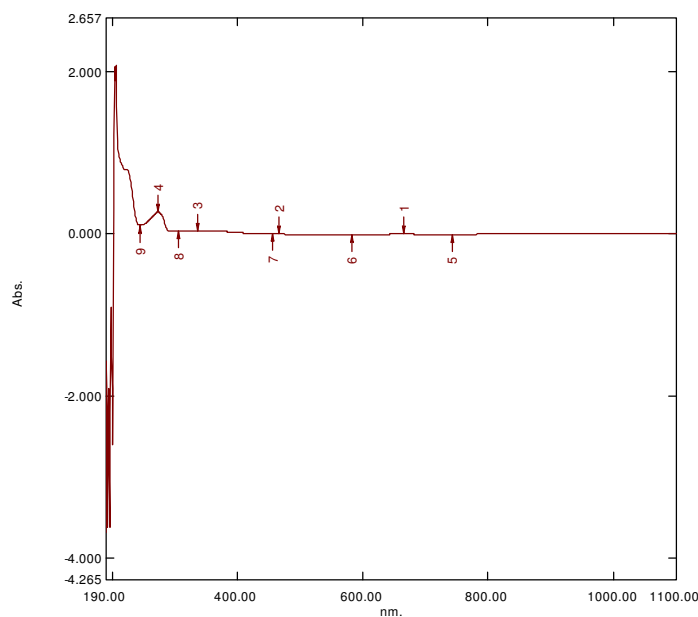


Fig. 1. Ultraviolet-Visible Spectroscopy analysis of ethanolic extracts of *Pavonia zeylanica*.

Table 3: FT-IR peak values and functional group of ethanolic extract of *Pavonia zeylanica*.

Extracts	Peak value	Functional group	Name of functional group	Vibrations
Ethanolic extracts of <i>Pavonia zeylanica</i>	688.59	C-H	Alkane	Stretch
	1043.49	C-F	Haloalkane	-
	1265.3	O-H	Hydroxyl	Bending
	1427.32	Carbonates	-	-
	1643.35	C-F	Haloalkane	-
	2362.8	C≡N	Nitrile	Stretch
	2929.87	O-H	Alkane	Stretch
	3398.57	N-H	Amine	Stretch

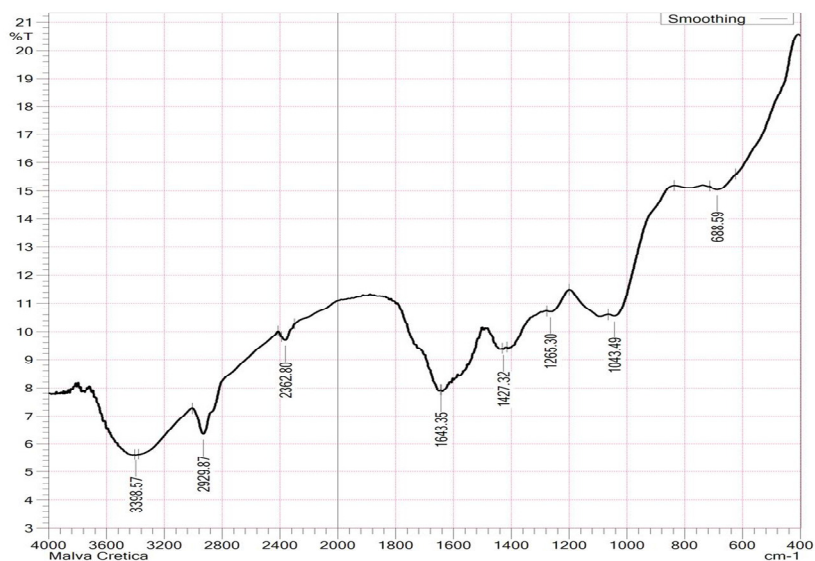


Fig. 2. Fourier-Transform Infrared Spectroscopy analysis of ethanolic extract of *Pavonia zeylanica*.

Table 4: GC-MS study of the ethanolic extract of *Pavonia zeylanica*.

Sr. No.	Retention time	Compound name	Molecular formula	Molecular weight	Area %	Molecular structure	Hit spectrum	Functional group	Biological activity
1.	2.528	Ethane, 1,1-Diethoxy	C ₆ H ₁₄ O ₂	118	7.376			Acetal	Antibacterial activity
2.	9.201	4h-Pyran-4-One, 2,3-Dihydro-3,5-Dihydroxy-6-Methyl	C ₆ H ₈ O ₄	144	5.161			Phenol	Antioxidant, analgesic, antipyretic and anti-inflammatory
3.	15.674	2-Methyl-6-Methylene-Octa-1,7-Dien-3-Ol	C ₁₀ H ₁₆ O	152	2.412			Methyl ester	Anti inflammation and antimicrobial activity
4.	16.344	3-Tetradecen-5-Yne, (E)	C ₁₄ H ₂₄	192	3.330			-	-
5.	16.719	Trans-Z-Alpha-Bisabolene Epoxide	C ₁₅ H ₂₄ O	220	1.797			Alcoholic compound	Antimicrobial activity
6.	18.445	3,7,11,15-Tetramethyl-2-Hexadecen-1-Ol	C ₂₀ H ₄₀ O	296	5.482			Methyl ester	Anti oxidant activity
7.	18.545	9-Methyl-Z-Z-10,12-Hexadecadien-1-Ol Acetate	C ₁₉ H ₃₄ O ₂	294	2.912			Methyl ester	Anti inflammation and antimicrobial activity
8.	19.790	N-Hexadecanoic Acid	C ₁₆ H ₃₂ O ₂	256	23.280			Saturated fatty acid	Anti microbial activity
9.	21.261	PHYTOL	C ₂₀ H ₄₀ O	296	6.412			Alcohol	Anxiolytic, metabolism-modulating, cytotoxic, antioxidant, autophagy and

									apoptosis – including antinociceptive, anti-inflammatory, immune – modulating and antimicrobial activity
10	21.481	1- Tridecyne	C ₁₃ H ₂₄	180	2.088			Hydroxyl	Anti microbial activity
11	21.546	3- Tetradecyn -1-Ol	C ₁₄ H ₂₆ O	210	11.767			Aromatic bicyclic	Anti cancerous activity compound
12	21.746	Pentadecanoic Acid	C ₁₅ H ₃₀ O ₂	242	5.291			Odd chain saturated fatty acid	-
13	23.552	OLEIC ACID	C ₁₈ H ₃₄ O ₂	282	2.984			Alkene	Anti microbial agents, transducer for immunosensor and its method of production, carcinogens, enzymes inhibitors
14	27.163	6,11-Dimethyl-2,6,10-Dodecatric n-1-Ol	C ₁₄ H ₂₄ O	208	2.711			Methyl	Liver detoxification, protein methylation, neurotransmitter synthesis, nucleic acid synthesis
15	30.060	Bisnorallocholic Acid	C ₂₂ H ₃₆ O ₂	332	2.613			Carboxylic acid	Anti microbial activity

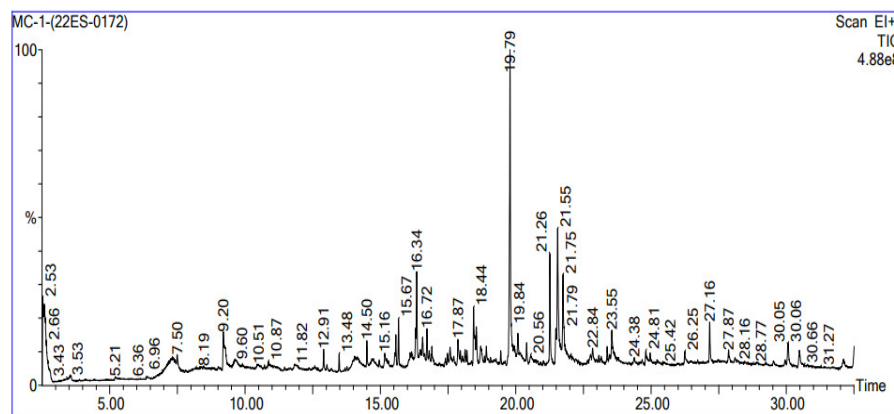


Fig. 3. GC-MS chromatogram of ethanolic extract of *Pavonia zeylanica*.

CONCLUSIONS

To conduct additional pharmacological analysis, the research plant *Pavonia zeylanica* may be exposed to the isolation of some more novel therapeutically useful compounds. This research has made it feasible to manufacture drugs for human consumption using these plants to treat a variety of illnesses. The plants also contained compounds that protected against oxidative stress. To expand the range of drugs now utilised in the treatment and prevention of human illnesses, more research is needed to investigate other bioactivities.

REFERENCES

- Anonymous (1962). The wealth of India: Raw Materials VI. Publications and Information Directorate, C.S.I.R., New Delhi.
- Abdullatif Azab (2017). Malva: Food, Medicine and Chemistry. *Eur. Chem. Bull.*, 6(7), 295-320.
- Ayaz, M., Sadiq, A., Junaid, M., Ullah, F., Subhan, F. and Ahmed, J. (2017). Neuroprotective and Anti-Aging Potentials of Essential Oils from Aromatic and Medicinal Plants. *Front. Aging Neurosci.*, 9, 168.
- Ayaz, M., Subhan, F., Ahmed, J., Khan, A.-U., Ullah F., Ullah I., Ali G., Syed N. I. H., and Hussain, S. (2015).

- Sertraline Enhances the Activity of Antimicrobial Agents against Pathogens of Clinical Relevance. *J. Biol. Res.*, 22, 4.
- Bano, I. and Deora, G. S. (2019). Preliminary Phytochemical Screening and GC-MS Analysis of Methanolic Leaf Extract of *Abutilon pannosum* (Forst. F.) Schlect. From Indian Thar Desert. *J. Pharmac. Phytochem.* 8(1), 894-899.
- Bokhad, M. N., Don, G. and Rothe, S. P. (2012). Preliminary phytochemical investigation of *Combretum albidum*. An ignored medicinally important liana. *J. Exp. Sci.*, 3(3), 1.
- Deshmukh, P. T., Fernandes, J., Atul, A. and Toppo, E. (2009). Wound Healing Activity of Calotropis Gigantea Root Bark in Rats. *J. Ethnopharmacol.* 125, 178-181.
- Dhivya, S. M. and Kalaichelvi, K. (2017). UV-Visible Spectroscopic and FT-IR analysis of *Sarcostemma brevistigma*, Wight. and Arn. *Int. J. of Curr. Pharmac. Res.*, 9(3), 46-49.
- Edeoga, H. O., Okwu, D. E. and Mbaebie, B. O. (2005). Phytochemical constituents of some Nigerian medicinal plants. *Afr. J. Biotech.*, 4(7), 685-688.
- Getie, M., Gebre-Mariam, T., Rietz, R., Neubert, R. H. H. (2002). Evaluation of the Release Profiles of Flavonoids from Topical Formulations of the Crude Extract of the Leaves of *Dodonea Viscosa* (Sapindaceae) *Die Pharmazie*, 57, 320-322.
- Griffiths, P. R. and de Haset, J. A. (1986). Fourier transforms infrared spectroscopy. New York: Wiley, 1986.
- Harborne, J. B. (1973). Phytochemical methods, Chapman and Hall, Ltd., London, 49-188.
- Harborne, J. B. (1998). Phytochemical methods. 3rded., London, Chapman and Hall.
- Hepcy Kalarani, D., Dinakar, A., and Senthilkumar, N. (2012). Antidiabetic, Analgesic and Anti-Inflammatory activity of Aqueous extracts of Stem and Leaves of *Alangium salvifolium* and *Pavonia zeylanica*. *Int. J. Drug Development & Res.*, 4(4), 298-306.
- Ivanova, D. G. and Singh, B. R. (2003). Nondestructive FTIR monitoring of leaf senescence and eliciting induced changes in plant leaves. *Biopolymers*, 72(2), 79-85.
- Jennings, W. and Shibamoto, T. (1982). Qualitative analysis of flavour and fragrance volatiles by glass capillary gas chromatography. *Mol. Nutri. Food Res.*, 26(9), 830.
- Joga Rao, Y. S. V. S., Annasamuel Lanka., Geetha Bhavani, K., Ramachandran, D., Bollikolla, H. B. (2020). Phytochemical Screening and GC-MS analysis of *Pavonia Zeylanica*. *Caribbean J. Sci. Tech.*, 8(1), 93-104.
- Komal Kumar, J. and Devi Prasad, A. G. (2011). Identification and comparison of biomolecules in medicinal plants of *Tephrosia tinctoria* and *Atylosia albicans* by using FT-IR. *Romanian J. Biophys.*, 21(1), 63-71.
- Lena, G. (2010). Phenolic compounds, antioxidant activity and in vitro inhibitory potential against key enzymes relevant for hyperglycemia and hypertension of commonly used medicinal plants, herbs and species in Latin America. *Bio. Resour. Technol.*, 10, 4676-4689.
- Leon Stephan Raj, T., Antony Selvi, A., Ramakrishnan, P., Antony Fency, M., Vellakani, M. and Vanila, D. 2015. Qualitative and quantitative analysis of phytoconstituents of *Micrococca mercurialis* (L.). Benth. *Ameri. J. Biol. Pharmac. Res.*, 2(4), 161-167.
- Lopes, G. C., Sanches, A. C. C., Nakamura, C. V., Dias Filho, B. P., Hernandez, L., and de Mello, J. C. P. (2005). Influence of Extracts of *Stryphnodendron polyphyllum* Mart. and *Stryphnodendron obovatum* Benth. on the Cicatrization of Cutaneous Wounds in Rats. *J. Ethnopharmacol.* 99, 265-272.
- Mehta, P., Shah, R., Lohidasan, S., and Mahadik, K. R. (2015). Pharmacokinetic Profile of Phytoconstituent (s) Isolated from Medicinal Plants—A Comprehensive Review. *J. Tradit. Complement. Med.*, 5, 207-227.
- Ofokansi, K. C., Esimone, C. O. and Anele, C. K. (2005). Evaluation of the *in vitro* combined anti-bacterial effects of the leaf extracts of *Bryophyllum pinnatum* (Fam. crassulaceae) and *Ocimum gratissimum* (Fam. Labiate). *Plant Prod. Res. J.*, 9, 23-27.
- Ovais, M., Ahmad, I., Khalil, A. T., Mukherjee, S., Javed, R., Ayaz, M., Raza, A. and Shinwari, Z. K. (2018). Wound Healing Applications of Biogenic Colloidal Silver and Gold Nanoparticles: Recent Trends and Future Prospects. *Appl. Microbiol. Biotechnol.*, 102, 4305-4318.
- Okwu, D. E. and Okwu, M. E. (2004). Chemical composition of *Spondias mombin* Linn. plant parts. *J. Sus. Agric. Environ.*, 6(2), 140-147.
- Perumal Samy, R., and Ignacimuthu, S. (2000). Antibacterial activity of some folklore medicinal plants used by tribals in Western Ghats of India. *J. of Ethnopharmac.* 69 (2000), 63-71.
- Prabhu, V. and Ramar, K. (2018). Phytochemical Screening by UV-VIS, FT-IR and GC-MS Spectroscopic Analysis of Leaf and Callus Extracts of Medicinal Plants *Sida schimperiana* Hochst. Ex A. Rich. *World J. of Pharmac. Res.*, 7(11), 728-743.
- Ridhambaradevi, G., Meenakshi Sundaram, M., Amala Hazel, A. M. and Banumathi, V. (2018). Physicochemical, Phytochemical, Bio-chemical analysis of Neuro Protective traditional Siddha medicine Chitramuttikudineer. *World J. of Pharm. Res.*, 7(18), 1195-1208.
- Shetty, N. P., Jørgensen, H. J. L., Jensen, J. D., Collinge, D. B. and Shetty, H. S. (2008). Roles of Reactive Oxygen Species in Interactions between Plants and Pathogens. *Eur. J. Plant Pathol.*, 121, 267-280.
- Shihabudeen, H. M. S., Priscilla, D. H. and Kavitha Thirumurugan (2010). Antimicrobial activity and phytochemical analysis of selected Indian folk medicinal plants. *Int. J. Pharma Sci. Res.*, 1(10), 430-434.
- Sodipo, O. A., Akini, J. A. and Ogunbamosu, J. U. (2000). Studies on certain characteristics of extracts of bark of *Pansynstali amacrueras* (K. Schemp) Pierre exbeille. *Global J. Pure Appl. Sci.*, 6, 83-87.
- Yang, J., and Yen, H.C.E. (2002). Early Salt Stress Effects on the Changes in Chemical Composition in Leaves of Ice Plant and Arabidopsis. A Fourier Transform Infrared Spectroscopy Study. *Plant Physiology*, 130, 1032-1042.
- Zohra, T., Ovais, M., Khalil, A. T., Qasim, M., Ayaz, M., and Shinwari, Z. K. (2019). Extraction Optimization, Total Phenolic, Flavonoid Contents, HPLC-DAD Analysis and Diverse Pharmacological Evaluations of *Dysphania ambrosioides* (L.) Mosyakin & Clemants. *Nat. Prod. Res.*, 33, 136-142.

How to cite this article: S.M. Dhivya, P. Vijayashalini, P. Abirami and S. Sharmila (2024). Scientific Investigation of *Pavonia zeylanica* (L.) Cav. A Potent Medicinal Plant. *Biological Forum – An International Journal*, 16(11): 67-74.